

Vol. 5 No. 2 (Feb 2012)

ISSN: 0974- 6846

Graft copolymerization of Gelatin-g-poly (Acrylic acid-co-Acrylamide) and calculation of grafting parameters

Fatemeh Soleimani¹, Mohammad Sadeghi^{2*}, and Hadis Shahsavari³ ^{1,3} Young researchers club, Islamic Azad University, Khorramabad Branch, Khorramabad,Iran ²Department of Chemistry, Science Faculty, Islamic Azad University, Arak Branch, Arak, Iran. m-sadeghi@iau-arak.ac.ir

Abstract

In this research, we synthesized a novel graft copolymer of gelatin-based via radical polymerization mixtures Acrylic acid (AcA) and Acrylamide (AAm) onto gelatin backbones. The polymerization reaction was carried out in an aqueous medium and in the presence of ammonium persulfate (APS) as an initiator. Evidence of grafting was obtained by comparing FTIR and TGA analysis of CMC and the graft copolymer as well as solubility characteristics of the products. The effect of grafting variables, i.e. concentration of AAm, AcA, APS and Gelatin, and temperature was systematically optimized to achieve a highest percent grafting possible. The overall activation energy for the grafting was also estimated to be 23.30kJ/mole.

Keywords: Gelatin, Graft copolymerization, Acrylamide, Acrylic acid, Optimization.

Introduction

Free radical vinyl graft copolymerization onto natural polymers is a well-known method for synthesis of natural-based superabsorbent hydrogels. In fact, an efficient approach to modify of natural polymers in order to synthesis of natural-based SAPs is graft polymerization of vinylic monomers onto their backbones in the presence of cross-linkers. Free radical graft copolymerization with various monomers can carried out with different initiator systems (Sandle et al., 1987; Athawale & Rathi, 1997; Heinze & Liebert, 2001). Although hydrogels made from synthetic polymers, such as polyacrylate posses excellent water-absorbing properties, their toxicity and non-bio-degradability might pose lona-time environmental problems and limit their use in drug delivery systems and consumer products. Natural-based SAPs have attracted much attention in medical and pharmaceutical fields because of their non-toxicity, biocompatibility and biodegradability (Athawale & Rathi, 1999).

As a protein, gelatin is a biomaterial with the above mentioned essential properties. Generally, crosslinking in gelatin is used in various purposes such as gelatin swelling and gelatin hydrogels as biodegradable implants to deliver small and macromolecular drugs. Recently, attention has been focused on employing gelatin substrate to produce hydrogels with a specific response to a biological environment (Hebeish & Guthrie, 1981; Okieimen & Ogbeifun, 1996; Mohammad Sadeghi & Nahid Ghasemi, 2012).

Although, APS-initiated grafting of vinyl monomers such as methyl acrylate, ethyl acrylate and ethyl methacrylate (Okieimen, 1998), AN/methyl methacrylate mixture (Okieimen & Ogbeifun, 1996), acrylamide (AAm) (Okieimen, 2003), and 4-vinylpyridine (Leza *et al.*, 1989) onto gelatin has been reported. However, to the best of our knowledge, no report has been published on the optimization graft polymerization of Acrylic acid (AcA) and Acrylamide (AAm) together onto Gelatin chains using APS-Protein initiating system. In the present study, to modify the Gelatin, the grafting of Acrylic acid and Acrylamide onto Gelatin chains in the presence of ammonium persulfate (APS) as an initiator was performed in a homogeneous system. The effect of reaction variables affecting on percent grafting was investigated.

Experimental

Materials

Gelatin (from Parvar Novin-E Tehran Co.), potassium persulfate (APS, from Fluka), Acrylamide (Merck) and Acrylic acid (Merck) were used without further purification. All other chemicals were also analytical grade. Double distilled water was used for graft copolymer preparation.

Preparation of Graft Copolymer

procedure for Α general chemically graft copolymerization of Acrylamide (AAm) and Acrylic acid (AcA) onto gelatin backbones was conducted as follows: Gelatin (1.0 g) was added to a three-neck reactor equipped with a mechanical stirrer (Heidolph RZR 2021, three blade propeller type, 300 rpm), including 35 mL double distilled water. The reactor was immersed in a thermo-stated water bath preset at a desired temperature (70°C). Then 0.10 g of APS as an initiator was added to gelatin solution and was allowed to stir for 10 min. After adding APS, variable amounts of AAm and AcA (AAm 0.50-2.50 g, AcA 0.50-2.50 g) were added simultaneously to the gelatin solution. After 90 min, the reaction product was allowed to cool to ambient temperature. The graft copolymer was poured to excess non solvent ethanol (200 mL) and remained for 3 h to dewater. Then ethanol was decanted and the product slices to small pieces (diameter \approx 5 mm). Again, 100 mL fresh ethanol was added and the graft copolymer was remained for 24 h (Pourjavadi & Zohuriaan-Mehr, 2002).

Homopolymer extraction

The graft copolymer, namely gelatin-g-Poly (AAm-co-AcA), was freed from homo-polymers, by pouring 1.00 g of the product in 75 mL of DMF solution. The mixture was stirred gently at room temperature for 48 h. After



Vol. 5 No. 2 (Feb 2012)

macroradicals

ISSN: 0974-6846

complete removal of the homo-polymers by under nitrogen atmosphere. Experiments were performed at a heating rate of 10°C/min until 700°C. centrifugation, the gelatin-g-Poly(AAm-co-AcA), was

Scheme 1. Proposed mechanistic pathway for synthesis of gelatin-g-poly(AAm-co-AcA) copolymer.



Gelatin backbone (R: various chains of 20 different amino acid)





filtered and washed with methanol and dried in oven at 50°C to reach a constant weight (Pourjavadi & Zohuriaan-Zohuriaan et al., 2005) Although, Mehr. 2002: dimethylformamide is a good solvent for PAAm and PAcA as well as a precipitant for pure gelatin or grafted gelatin, so the homo-polymers could be easily separated from the rough products. However, it seemed to be difficult to further separate the unreacted gelatin from the products and the right separation methods are still in progress in my research. In view of the modification intention, the unreacted gelatin is not very necessary to be separated from the products. So the blends of unreacted gelatin and the graft copolymer CMC-q-Poly (AAm-co-PAcA) were actually obtained in this research and their compositions were unknown. In several studies on the grafting modification of polymer, the unreacted substrate polymer and graft copolymer were also not separated from the products (Zhang et al, 2000; Zhang, 2000; Ibrahim et al., 2002). If the unreacted gelatin could be separated from the products, the graft copolymer CMC-g-Poly(AAm-co-PAcA) with higher G% could be obtained, but the values of PC% and GE% would not be affected. Instrumental analysis

also performed on a Universal V4.1D TA Instruments (SDT Q600) with 8-10 mg samples on a platinum pan

gelatin-g-Poly(AAm-co-AcA) samples The characterized as KBr pellets using a Mattson-1000 FTIR spectrophotometer. Thermo-gravimetric analyses were Grafting parameters

The grafting parameters, i.e. grafting-rati (Gr %), grafting-efficiency (Ge %), add-on value (Ad %), and homopolymers content (Hp %) used to characterize the nature of the copolymer are defined and calculated using the following equations (Fanta & Doane, 1986):

$Gr \% = 100 (W_2 - W_0) / W_0$	(1)
Ad %= 100 ($W_2 - W_0$) / W_2	(2)
$\Box_{n} 0/ 100 (M/ M/ M$	(2)

Hp % = 100 ($W_1 - W_2$) / W_1 (3)

Ge $\% = 100 (W_2) / W_1$ (4)

where W_{0} , W_{1} , and W_{2} are the weight of the initial substrate, total product (copolymer and homopolymers) and pure graft copolymer (after DMF extraction) respectively.

Results and discussion

Synthesis and spectral characterization

The mixture of monomers, acryl-amide and acrylic simultaneously grafted onto gelatin acid was backbones in a homogeneous medium using APS as a radical initiator. A general reaction mechanism for gelatin-g- poly(AcA-co-AAm) copolymer formation is shown in Scheme 1. At the first step, the thermally dissociating initiator, i.e. APS, is decomposed under heating to produce sulfate anion-radical. Then, the anion-radical abstracts hydrogen from one of the functional groups in side chains (i.e. COOH, SH, OH, and NH₂) of the substrate to form corresponding radical. So, these macroradicals initiated monomers

were



Vol. 5 No. 2 (Feb 2012)

2043

ISSN: 0974-6846

grafting onto gelatin backbones led to a graft copolymer (Mohammad & Hossein, 2010).

Infrared spectroscopy was carried out to confirm the chemical structure of the copolymer. Fig.1 shows the FTIR spectra of the gelatin substrate and the synthesized copolymer. The band observed at 1634 cm⁻¹ can be attributed to C=O stretching in carboxamide functional groups of substrate backbone (Fig.1a). The graft copolymer product comprises a gelatin backbone with side chains that carry carboxylate and carboxamide functional groups that are evidenced by peaks at 1558 and 1637 cm⁻¹ respectively (Fig. 1b). The characteristic band at 1558 cm⁻¹ is due to asymmetric stretching in carboxylate anion that is reconfirmed by another peak at 1411 cm⁻¹ which is related to the symmetric stretching mode of the carboxylate anion (Zhang et al., 2000; Ibrahim et al., 2002). The stretching band of the grafted carboxamide groups overlapped with that of the gelatin portion of the copolymer.

Thermogravimetric behavior

The grafting was also supported by thermo-gravimetric analysis (Fig. 2). TGA of gelatin (Fig. 2a) shows a weight loss in two distinct stages. In the TGA curve gelatin-gpoly (AAm-co-NaAcA) copolymer about 10-12% loss in weight is observed below 130°C. This was attributed to the removal of the absorbed water. Also the Fig.2 shows that the degradation of native gelatin is faster than that of grafted gelatin. About 45% weight loss takes place in the temperature of 280°C for gelatin. A residual weight of 72% is observed at 280°C for gelatin-g-poly (AAm-co-NaAcA) copolymer. In general, the copolymer had lower weight loss than Gelatin. This means that the grafting of Gelatin increases the thermal stability of Gelatin in some extent (Doyle, 1961).

Solubility test

To obtain an additional evidence of grafting, solubility difference between the grafted and the non-grafted polymer was used. Gelatin and poly (AAm-co-AcA) are soluble in water and DMF respectively. When a reaction product was extracted with DMF and alternatively with water for 48 h, an insoluble solid still remained. A physical mixture of gelatin and poly (AAm-co-AcA) was treated in the same way and was found to dissolve completely. Therefore, it is obvious that the resulted graft copolymer was not a simple physical mixture, but some chemical bonds must exist between the gelatin substrate and poly (AAm-co-AcA) macromolecules (Mohammad & Hossein, 2010).

Optimization of polymerization

In the present investigation, the effect of concentration of gelatin, APS, AcA and AAm, along with reaction temperature was studied to optimize the reaction conditions. It may be found from the related curves (next figures) that the trends of the "changes" are similar for grafting parameters Gr, Ge, Hp, and Ad. The reason is the similar concepts applied for defining the grafting parameters (Eqs. 1-4).

Effect of initiator concentration

effect of concentration APS on The graft polymerization was studied by changing its concentration from 0.0003 to 0.0008mol/L (Fig. 3). It was observed that the grafting percent is increased versus increasing the APS concentration from 0.01 up to 0.00055 mol/L and then, it is decreased considerably with a further increase in the amount of APS. The maximum grafting percent (160%) is obtained at APS 0.00055 mol/L. The number of active free radicals on the gelatin backbone is increased in terms of the initiator levels lower than 0.00055 mol/L. This accounts for the initial increment in grafting percent up to a certain amount of APS. The grafting percent decrease after the maximum may be attributed to increased number of produced radicals led to terminating step via bimolecular collision resulting in enhanced crosslink density (Pourjavadi & Zohuriaan-Mehr, 2002).

An additional reason for decreasing the grafting percent can be related to decreasing molecular weight (MW) of the grafted polyacrylamide and polyacrylic acid at high levels of APS concentration. Since MW inversely depends on initiator concentration (I), and the higher I results in lower MW in turns to be a lower grafting percent of the copolymer (Zohuriaan et al., 2005). On the other hand, free radical degradation of gelatin substrate is also possible at high APS levels. A similar observation is recently reported in the case of degradation of chitosan with potassium persulfate (Hsu & Don, 2002).

Effect of reaction temperature

To study the influence of the reaction bath temperature on the grafting parameters, the grafting of AAm, AcA onto gelatin was carried out at temperature ranging from 35-70°C. As shown in Fig. 4, higher temperatures favor the rate of diffusion of monomers to the gelatin macroradicals as well as increase the kinetic energy of radical centers. In addition, higher temperatures increase the rate of decomposition of the thermally dissociating initiator APS (Zohuriaan et al., 2005). The temperatures higher than the optimum value (55°C), however, lead to low-graft copolymer. This percent-loss may be attributed to (a) oxidative degradation of gelatin chains by sulfate radical-anions, (b) increasing the rate of termination and chain transfer reactions, and (c) decomposition of APS to give O_2 (a radical scavenger), which reacts with primary free radicals (Eqs. 5 and 6), (Hsu et al., 2002), resulting in decreased molecular weight and decreased grafting (the sulfate radical anions may react with water to produce hydroxyl radicals (Eq. 5) and finally oxygen (Eq. 6)).

SO, + H,O —→ HSO, + HÓ (5)

The rates graft copolymerization (R_{a}) may be evaluated as measures of the rate of monomer disappearance by using the following equations (Fanta & Doane, 1986):

(6)



Vol. 5 No. 2 (Feb 2012)

ISSN: 0974-6846

Fig.1. FTIR spectra of pure gelatin (a) and gelatin-g-poly (AAm-co-NaAcA) copolymer(b).



Fig. 2. TGA curves of (a) gelatin and (b) gelatin-g-Poly (AAm-co-AcA).



Fig.3. Grafting percent variances with concentration of ammonium persulfate variance



Fig. 4. Grafting percent variances with temperature variance







1/T (K⁻¹)

Fig. 6. Grafting percent variances with amount of acrylamide monomer variance



 $Rg(mol. s^{-1}.m^{-3})$

Weight of grafted polymer

 $\frac{1}{Molecularweight of monomer \times [rection time (s)] \times volume (m^3)}$ (7)

Overall activation energy of grafting (E_a) may also be estimated from the temperature data through plotting lnR_g versus 1/T ($^{o}K^{-1}$) for the initial portion of the data of the temperature series given in above text. The slope of this Arrhenius plot (Fig. 5) resulted in a rough estimation of Ea of grafting using the relationship slope = $-E_a/R$; where R is the universal gas constant. Therefore, Ea for the graft copolymerization was found to be 23.30 kJ/mole (5.57 kcal/mole).

Effect of AAm concentration

The effect of monomer amount on the grafting reaction was studied at various concentrations of AAm while other influential factors were unchanged. The grafting parameter variations are changed by the amount of

Fig. 7. Grafting percent variances with amount of AMPS monomer variance



Fig. 8. Grafting percent variances with amount of gelatin variance.





Vol. 5 No. 2 (Feb 2012) ISSN: 0974- 6846

charged monomer (Pourjavadi & Zohuriaan-Mehr, 2002). The results are given in Fig. 6. The grafting extent is significantly increased due to more availability of monomer for grafting. However, beyond a certain Gr value, *i.e.*, 142% at AAm 3.5 mL, the trend is inversed. The conversion and the grafting efficiency (Ge) are decreased, and homopolymer content is increased noticeably from 4 to 14 percent. Thus, acrylamid in an amount of AAm 3.5 mL was recognized as an optimum monomer concentration. Once the monomer units are added, an excess of monomer can only increase the optimum volume of the reaction mixture (Pourjavadi & Zohuriaan-Mehr, 2002; Zohuriaan, 2005).

Effect of AcA concentration

The Acrylic acid concentration was varied from 0.45 to 1.30 mol/L equal (1.5-4.7 mL) to study its effects on grafting parameters (Fig. 7). These parameters were found to be increased by enhancement of AcA concentration from 0.45 up to 0.84 mol/L. This behavior can be attributed to the increase of monomer concentration in the vicinity of the gelatin backbone and consequent greater availability and enhancement chances for molecular collisions of the reactants. The decrease in %Gr and %Ad after a certain level of AcA (0.84 mol/L) is probably due to preferential homopolymerization over graft copolymerization as well as increasing the viscosity of reaction medium, which hinders the movement of free radicals(Mohammad & Hossein, 2010). Needless to say, the increase in the chain transfer to monomer molecules may be other possible reason for the diminished grafting at higher AcA concentrations. Similar observations have been reported for the grafting of ethyl acrylate onto cellulose (Okieimen & Ogbeifun, 1996; Ibrahim et al., 2002) and methyl acrylate onto starch (Athawale & Lele, 2002). Effect of gelatin concentration

The related to the grafting dependence on Gelatin amount is summarized in Fig.8. Maximum grafting percent and the lowest homopolymers formation was observed at 0.80 g Gelatin, while others reactants including, monomers, initiator, and temperature were kept constant. Beyond this value, both grafting percent and add-on values are considerably reduced. This behavior is attributed to the availability of more grafting sites for initiation of graft copolymerization at higher concentration of the substrate (until 0.80 g Gelatin). However, upon further increase in the substrate concentration, increase in the reaction medium viscosity restricts the movements of macroradicals leading to decreased grafting percent and add-on values (Hsu et al., 2002). It also may be attributed to deactivation of the macroradical growing chains (e.g., by transfer reactions, combination and/or interaction with the primary radicals) soon after their formation (Mohammad & Hossein, 2010).

Conclusion

The monomers, Acrylamide (AAm) and Acrylic acid (AcA) can be easily graft copolymerized onto gelatin

"A novel graft co-polymer" http://www.indjst.org



References

- 1. Athawale VD and Lele V (2000) Thermal studies on granular maize starch and its graft copolymers with vinyl monomers. Starch/Starke. 52, 205-213.
- 2. Athawale VD and Rathi SC (1997) Role and Relevance of polarity and solubility of vinyl monomers in graft polymerization onto starch. React. Func. Polym. 34, 11-17.
- 3. Athawale VD and Rathi SC (1999) Graft polymerization: Starch as a model substrate. J. Macromol. Sci. Rev. Macromol. Chem. Phys. C39(3), 445-480.
- 4. Deshmukh SR, Sudhakar K aand Singh RP (1991) Drag-Reduction efficiency, shear stability and biodegradation resistance of carboxymethyl cellulose-based and starch-Based copolymers. J. Appl. Polym. Sci. 43, 1091-1101.
- 5. Doyle CD (1961) Estimating thermal stability of experimental polymers by empirical thermogravimetric analysis. Anal. Chem. 33(1), 77-79.
- 6. Fanta GF and Doane WM (1986) Grafted starches. In: Modified starches: Properties and uses; Wurzburg OB (Ed.), CRC Press: Boca Raton (Florida). pp: 149-178.
- 7. Flefel EM, Ibrahim MM, El-Zawawy WK and Ali AM (2002) Graft copolymerization of N-vinylpyrrolidone and Acrylamide on cellulose derivatives: Synthesis, Characterization and study of their biological effect. Polym. Adv. Technol. 13, 541-547.
- 8. Hebeish A and Guthrie JT (1981) The chemistry and technology of cellulosic copolymers. Springer, Berlin.
- 9. Heinze T and Liebert T (2001) Unconventional methods in cellulose functionalization. Prog. Polym. Sci. 26, 1689-1762.
- 10. Hsu SC, Don TM and Chiu WY (2002) Free radical degradation of chitosan with potassium persulfate. Polym. Degrad. Stab. pp: 75-73.
- 11. Ibrahim MM, Flefel EM and El-Zawawy WK (2002) Cellulose membranes grafted with vinyl monomers in homogeneous system. J. Appl. Polym. Sci. 84, 2629-2638.



Vol. 5 No. 2 (Feb 2012)

ISSN: 0974-6846

- 12. Leza ML, Casinos I and Guzman GM (1989) Graft copolymerization of 4-vinylpyridine onto cellulosics: Effect of temperature. Eur. Polym. J. 25(12), 1193-1196.
- 13. Leza ML, Casinos I and Guzman GM (1990) Graft copolymerization of 4-vinylpyridine onto cellulosics: Effects of stirring and Inorganic salts. Brit. Polym. J. 23, 341-346.
- 14. Mohammad S and Hossein H (2010) Studies on graft copolymerization of 2-hydroxyethylmethacrylate onto kappa-carrageenan initiated by ceric ammonium nitrate. Journal of The Chilean Chemical Society. 55,N⁰ 4. doi: 10.4067/S0717-97072010000400019
- 15. Mohammad Sadeghi and Nahid Ghasemi (2012) Preparation and swelling behavior of carrageenan-graftpolymethacrylamide superabsorbent hydrogel as a releasing drug system. Indian J.Sci.Technol. 5 (2), this issue.
- 16. Okieimen FE (1998) Graft copolymerization of vinyl monomers on cellulosic materials. Angew. Makromol. Chem. 260(1), 5-10.
- 17. Okieimen FE (2003) Preparation, characterization and properties of cellulose-polyacrylamide graft copolymers. J. Appl. Polym. Sci. 89, 913-923.
- 18. Okieimen FE and Ogbeifun DE (1996) Graft copolymerizations of modified cellulose: Grafting of methyl acrylate ethyl acrylate and ethyl methacrylate on carboxymethyl cellulose. Eur. Polym. J. 32(3), 311-315.
- 19. Okieimen FE and Ogbeifun DE (1996) Graft copolymerizations of modified cellulose: Grafting of acrylonitrile and methyl methacrylate on carboxymethyl cellulose. J. Appl. Polym. Sci. 59, 981-986.
- 20. Pourjavadi A and Zohuriaan-Mehr MJ (2002) Modification of carbohydrate polymers via grafting in air. 1. Ceric-Induced synthesis of starch-g-polyacrylonitrile in presence and absence of oxygen. Starch/Starke. 54, 140-147.
- 21. Sandle NK, Verma OPS and Varma IK (1987) Thermal characterization of starch-g-acrylonitrile copolymers. Thermochim. Acta. 115, 189-198.
- 22. Zhang J, Zhang LM and Li ZM (2000) Synthesis and aqueous solution properties of hydrophobically modified graft copolymer of sodium carboxymethyl cellulose with acrylamide and dimethyloctyl (2-methacryloxyethyl) ammonium Bromide. J. Appl. Polym. Sci. 78, 537-542.
- (2000) 23. Zhang LM Modification of sodium carboxymethylcellulose grafting by of diallyldimethylammonium chloride. Macromol. Mater. Engg. 280/281, 66-70.
- 24. Zhang LM and Tan YB (2000) Graft copolymerization of 2-(Dimethylamino)ethyl methacrylate onto Carboxymethylated Cellulose. Macromol. Mater. Engg. 280/281, 59-65.
- 25. Zohuriaan MJ, Pourjavadi A and Sadeghi M (2005) Modified CMC. Optimized synthesis 1. of Carboxymethylcellulose-g-Polyacrylonitrile. Iranian Polymer J. 14(2), 131-138.