MS 08113

Journal of Pharmaceutical Research Vol. 8, No. 3, July 2009 : 167-169.

EVALUATION OF ANTI-HEPATOTOXIC ACTIVITY OF *CLERODENDRUM PHLOMIDIS* L. ON CARBON TETRACHLORIDE INDUCED HEPATIC INJURY IN RATS

Ahmed B¹, Verma A¹ and Masoodi MH²*

¹Department of Pharmaceutical Chemistry, Faculty of Pharmacy, Jamia Hamdard, Hamdard Nagar, New Delhi - 110062, India.

²Department of Pharmaceutical Sciences, Kashmir University, Hazratbal, Srinagar, J & K - 190006, India.

Received on: 28.07.2008	Revised : 22.07.09	Accepted : 30.07.09
-------------------------	--------------------	---------------------

ABSTRACT

Methanol extract of the stem of *Clerodendrum phlomidis* L. was investigated for anti-hepatotoxic activity against CCl₄ induced hepatic damage in male Wister rats. The extracts in dose of 500mg/kg b.w p.o for 7 days were compared with the standard silymarin (Silybon–70, 10mg/kg-b.w p.o). The methanol extract of the plant have shown anti-hepatotoxic activity by reducing the elevated levels of serum enzymes such as serum glutamate oxaloacetate transaminase (SGOT) by 29.88 %, serum glutamate pyruvate transaminase (SGPT) by 25.86 % while alkaline phosphatase (ALP) 14.21 %. On the other hand, total protein (TP) were increased by 27.20 %, as compared to standard drug silymarin which decreased SGOT by 49.66 %, SGPT by 50.31 %, ALKP by 33.29 % and increased TP levels by 49.40 %. These biochemical observations were also supplemented by histopathological examinations of the liver sections

Keywords: Antihepatotoxic; Carbon tetrachloride; Clerodendrum phlomidis; Silymarin.

INTRODUCTION

Herbal medicines have been used in the form of vegetables, drugs or their extracts for the treatment of diseases and maintaining health. Various commercial preparations available from the crude plant extracts are available as formulations for the treatment of liver ailments¹. However, exposure to various therapeutic agents and environmental pollution leads to various disorders of organs, especially of liver². Clerodendrum phlomidis L. (Verbenaceae), commonly known as 'Arni' a shrub 0.9-2.4 m. high, scarcely woody, not much branched; stems bluntly quadrangular; young parts usually glabrous, leaves often ternate as well as opposite, oblong. The plant has been used in dropsy³. The decoction of roots is slightly aromatic and astringent and is used as a demulcent in gonorrhea. It is also given to children during convalescence of measles. The juice of leaves is used as bitter tonic⁴ and also given in neglected syphilitic complaints5. Psychopharmacological activity⁶, antidiarrhoeal activity7, antimutagenic activity8 and antifungal activity9 are some of its reported biological activities. The present article reports the anti-hepatotoxic activity of methanol extract. The ethyl acetate and hexane extracts of leaves and stems of C. phlomidis showed antifungal activity against plant and human pathogens but it is more effective in plants. It was tested by poison plate technique9.

EXPERIMENTAL Plant Material

The stem of *Clerodendrum phlomidis* were obtained from the market of Khari Bavli, Chandi Chauk, Old Delhi. A voucher specimen (CP-FP-32) of the plant has been kept in the herbarium of Jamia Hamdard University for further reference.

Preparation of plant extract

The plant material (1.5 kg) was dried and crushed to coarse powder and extracted with ethanol using cold percolation method till completely exhausted. The ethanol extract was then dried under reduced pressure to get the crude dried fraction of methanol 120.0 gm.

Experimental animals

Male Albino Wistar rats weighing 150–200 gm were employed for assessing the antihepatotoxic activity. They were procured from the Central Animal House of Jamia Hamdard, New Delhi (173/CPCSEA), after approval under the project proposal number-326. They were fed with a standard pellet diet and water *ad libitium*. The animals were maintained at 25 to 28 °C with 40-70% RH and 12 h light/dark cycles and were fasted for 12 hours prior to the experiment.

Antihepatotoxic activity

The animals were divided into four groups consisting (5 each). The first group (I) served as normal control which received normal saline only. The second group (II) served as toxic control and received CCI_4 diluted

*Correspondence : mube5@yahoo.com, mubashir@kashmiruniversity.ac.in, M : +91 9419076525

Journal of Pharmaceutical Research Vol. 8, No. 3, July 2009 : 167

with liquid paraffin in a ratio of (1:1) (1.5 ml/kg b.w, i.p.) on the first day to produce toxicity in the liver¹⁰. The third group (III) was given a single dose of CCI, on the first day (1.5 ml/kg b.w, i.p.) and then silymarin (Slybon-70, 10 mg/kg b.w, p.o.) was given for 7 days in the form of suspension using 1% Tween - 80). Group (IV) received a single dose of CCI, on the first day (1.5 ml/ kg b.w, i.p.) and then methanol extract at the dose of (500 mg/kg b.w, p.o.) for 7 days in the form of suspension using 1% Tween - 80)11. On the day 8 the blood samples were withdrawn by puncturing the orbital plexus. The blood samples were allowed to clot for 30-40 min at room temperature. Serum was separated by centrifugation at 2500 rpm for 15 min. Serum from 5 animals was taken for biochemical evaluation. Two rats from each group were sacrificed by decapitation and their livers were studied for histopathology.

Assessment of liver function

Various biochemical parameters like serum SGOT, SGPT¹², ALP¹³, and TP¹⁴ were carried out by reported methods.

Statistical analysis

The data of biochemical estimations were reported as mean of the data standard error (S.E), where n = 5. For determining the statistical significance one way analysis of variance (ANOVA) and Dunnett's test was employed. P–values of less than 0.05 were considered significant¹⁵.

Histopathological studies

For histopathological study, the livers were quickly removed after autopsy and fixed in 10% formalin¹⁶. The rats were sacrificed and the livers removed were washed with normal saline. Small pieces of tissues were embedded in paraffin wax. The sections of about 5-6 m were cut, stained and then observed under microscope for histopathological changes in liver and their pictographs were taken.

RESULTS AND DISCUSSION

Results in Table-1 show that the animals of group (II), who received only CCl, were found to develop significant hepatic damage as was observed from elevated levels of SGOT, SGPT and ALP and decrease in TP levels as compared to group (I) normal animals. However, in group (IV) treated with methanol extract of C. phlomidis significantly reduced CCl, induced elevation of liver enzymes such as SGOT by 98.62 units/ml, SGPT by 95.20 units/ml, while ALP by 60.04 units/ml respectively and TP level was increased by 5.33 gm/dl respectively. Standard drug silymarin (Slybon) also decreased SGOT by 70.80 units/ml, SGPT by 63.80 units/ml, ALKP by 46.69 units/ml and increased TP levels by 6.26 gm/dl against CCl, intoxicated rats. The above results indicated that the methanol extract of C. phlomidis was possessing antihepatotoxic activity.

Ahmed B, Verma A and Masoodi MH

Table 1: Effect of methanol fraction of Clerodendrum phlomidis stem on serum enzymatic activity in CCl₄ induced liver damage in rats.

Group s n=6	Treatment	Do ce	SGOT unitsimi	SGPT unitsmi	ALKP unitsimi	TP Gm/di
1	Normal canitol		5+5+± 1.25"	44.45± 1.36"	43.49± 1.73**	6.29±0.40**
а	Tate cont d	1.5 milkg (1.p.)	140.65± 1.81	128.41±2.48	69.99± 2.21	4.19±0.38
<u>а</u> ш.:	Shmatin (Slandard drug)	10 mg/kg (p.o)	7080± 1.62"	6380± 209"	45.69± 1.12"	6.25±0.45**
IV	Me Iranol extract	SDOmgikg (p.c.)	9862± 201"	95.20± 1.79"	60.04± 1.80'	5.33±0.15**

SGOT, serum glutamyl oxaloacetate transaminase; SGPT, serum glutamate pyruvate transaminase; ALKP, alkaline phosphate: TP, total protein; i.p. intraperitonally; p.o. per oral. ** P < 0.01; *P < 0.05 vs CCl₄. Values are mean ± S.E. of five animals. One way analysis and Dunnett's test.

Similarly histopathological studies in Table 2 reveal that in animals of group (I) normal control, liver samples show normal architecture (Fig.1). In toxic control group (II), CCI, caused fatty deposition and necrosis of hepatocytes (Fig.2). In group (III) treated with standard drug silymarin, liver samples showed a good recovery with absence of necrosis and fatty depositions (Fig.3). In group (IV) treated with methanol extract of C. phlomidis showed a significant recovery of hepatocytes and liver histology was almost normal in them (Fig.4), which were in accordance with the results obtained from biochemical parameters (Table 1). The results thus indicate that methanol extract of C. phlomidis (stem) possessed antihepatotoxic activity. Further work needs to be carried out to isolate the active principle responsible for antihepatotoxic activity.

	Table	2:	Histopathological	changes
--	-------	----	-------------------	---------

Group c I	Treatment Normal control	Microscopio observations Liter samples show normal archilecture without any degeneration, necrosisor inflammation.			
Ш	Toxic control (C Ci.)	Prominenti centrilobular faily change with prominentiand enlarged central vein along with significant perportal information reflecting liver diamage .			
ш	Slymarin (Slandard drug)	Samples showed a significant reduction in portal infammation and in the sinusoidal dialation. The central web was clearly utsible. Uver samples also showed good recovery with absence of necrosis and faily depositions.			
VI	CCI ₄ + Melhanol extract	Liver his lology was almost norm at with only very fille shousoidal dialation seen in some hepatic lobules. Central vein appeared clear with he disappearance of necrosis.			

Histopathology Slides



Fig. 1 Group I. High power photomicrograph of normal control rat liver on 8^{th} day (HE x 40X).

Journal of Pharmaceutical Research Vol. 8, No. 3, July 2009 : 168



Fig. 2 Group II. High power photomicrograph of toxic control rat liver on 8^{th} day (HE x 40X).



Fig. 3 Group III. High power photomicrograph of silymarin treated liver on 8th day (HE x 40X).



Fig. 4 Group IV. High power photomicrograph of Clerodendrum phlomidis methanol fraction treated rat liver on 8^{th} day (HE x 40X).

Ahmed B, Verma A and Masoodi MH

References

- 1. Handa SS, et al. Fitoterapia. 1986; 57: 307.
- 2. Ravinder K, et al. Ind J Expt Biol. 1994; 32: 328.
- Kirtikar KR, Basu BD. Indian Medicinal Plants. Dehradun: International Book Distributers 1999, p1947.
- Nandkarni AK. Dr. KM Nandkarni s Indian Materia Medica. Bombay : Popular Prakashan 2002, p 353.
- Chopra RN, et al. Glossary of Indian Medicinal Plants: (Council of Scientific and Industrial Research), New Delhi, 1968. p71.
- 6. Murugesan T, et al. Phytomedicine. 2001; 8: 472.
- 7. Rani S, et al. J Ethanopharmacol. 1999; 68: 315.
- 8. Reid KA, et al. J Ethnopharmacol. 2006; 106: 44.
- 9. Anita R, et al. Turk L Biol. 2006; 30: 139.
- Vogel HJ. In Drug Discovery and Evaluation, Pharmacological Assays, 2nd Edn. New York : Springer Verlag, Berlin Heidelberg, 2002, p924.
- 11. Masoodi MH, et al. J Pharmaceutical Research. 2007; 6: 190.
- 12. Reitzmann S, et al. Am J Clin Path. 1957; 28: 56.
- 13. Kind PRN, et al. J Clin Path. 1954; 7: 332.
- Wooton IDP, In Micro-analysis in Medical Biochemistry, 4th ed. London : J. and A. Churchill Ltd. 1964, p138.
- 15. Dunnet CW, Biometrics. 1964; 20: 482.
- Luna LG, In Manual of Histology, Staining methods of Armed Forces Institute of Pathology. 3rd Edn. New York : Mc Graw-Hill Book Co. 1986, p146.

Corrigendum			
Vol.8, No.2 A	pril 2009		
On page	For	Read	
105	Received on : 21.10.2009	21.10.2008	
108	Received on : 14.05.2009	02.12.2008	
	Revised : 21.05.09	21.03.09	
	Accepted : 22.05.09	22.04.09	