Fine mapping of consistent quantitative trait loci for yield under drought stress using rice (*Oryza sativa*) recombinant inbred lines adapted to rainfed environment

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Drought stress is a serious constraint, especially in rainfed rice production, and breeding for drought tolerance by selection based on yield under stress, though effective, is slow Mapping quantitative trait loci (QTLs) for yield and its components under drought stress predominant in rainfed target populations of environment (TPE) will help overcome this limitation. In the present study, a subset of 143 F₈ and F₉ recombinant inbred (RI) lines derived from IR62266-42-6-2 (IR62266), a high-yielding indica ecotype and Norungan, a landrace from TPE, was used to map QTLs for yield and its components under drought predominant in TPE. A large effect yield QTL observed under drought stress in TPE was consistent across two years with a phenotypic variation of 31.3% and 37.9% and additive effect of 629.2 and 424.9 kg/ha Further, this region was fine-mapped to 94.0 kb with positive effect on grain yield under stress.

Keywords: Comparative genomics, drought stress, fine mapping, quantitative trait locus, rice.

RICE (Oryza sativa L.) is one of the world's primary staple foods with 730 million tonnes (Mt) harvested in 2012 (ref. 1). It was cultivated in 163 million hectares (M ha) worldwide under varying ecosystems in 2013 (ref. 2). Globally 50% of the rice acreage is rainfed³. More than 23 M ha of rice is cultivated under rainfed environment in India⁴. In recent years, the increased occurrence and severity of drought stress has led to a yield decline in rice⁵. In rainfed regions, drought is the major environmental factor that reduces productivity by 13-35% (ref. 6). Developing cultivars with inherent capacity to tolerate drought stress will help improve rainfed rice production⁷. Direct selection for grain yield is an effective approach for the development of drought-tolerant varieties⁸⁻¹⁰. However, conventional breeding for drought resistance is slow in attaining progress due to difficulty in phenotyping large progenies and year-to-year variation in rainfall tolerance, quantitative trait loci (QTL) mapping followed by marker-assisted breeding (MAB) is considered as an alternate strategy to overcome this limitation¹¹. Though several QTLs for yield under drought stress have been mapped in rice¹²⁻¹⁶, only few have been successfully deployed in MAB till date¹⁷. Identifying consistent QTLs for yield under stress with large effect and fine-mapping them will speed up MAB for drought tolerance in rice^{12,18}. QTLs with large intervals have the chance of getting undesirably linked genes; hence it is important to fine-map the QTLs so as to introgress the precise genomic segment of interest. Thus, QTLs for yield under drought in upland condition have been fine-mapped recently¹⁹. However, drought stress differs markedly between upland and rainfed lowlands due to climate and inherent soil characteristics. Thus mapping QTLs under drought predominant in target populations of the environment (TPE) using population derived from locally adapted rice lines is critical^{20,21} to further improve the efficacy of MAB. Utilizing unexploited yield-related variabilities and QTLs from new cross combinations, especially involving wild species²² and landraces is important. In this study, we report identification of consistent,

pattern. Hence, for quantitative traits such as drought

In this study, we report identification of consistent, large-effect QTL for yield under drought stress in TPE, and fine-mapping of the qDTY6.1 region to 94.0 kb using local rice lines adapted to TPE with potential to develop high-yielding, drought-tolerant rice cultivars through MAB.

Materials and methods

Mapping population

Norungan, an *indica* landrace from Tamil Nadu, India, is drought-resistant and possess deep and thick roots with superior hardpan penetrating ability²³. IR62266, a lowland *indica* ecotype, has shallow and thin root system²³, but has the capacity for osmotic adjustment^{24,25}. A subset of 143 F₈ and F₉ RI lines derived from these two parents

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 $(IR62266 \times Norungan)$ was used in the present study for mapping QTLs for phenology, physio-morpholgical and yield traits under drought in TPE and further fine-map the consistent large-effect QTL for yield under drought.

Field experiments

The drought experiments were conducted in TPE during rainfed season (September-January) of 2011-12 (DS01) and 2012-13 (DS02) in the experimental fields of Agricultural Research Station, Tamil Nadu Agricultural University (TNAU), Paramakudi, India and during May-October 2013 (FC01) under irrigated (non-stress) condition in Paddy Breeding Station, TNAU, Coimbatore, India. In DS01 and DS02, a subset of 143 F₈ and F₉ recombinant inbred (RI) lines of IR62266 × Norungan respectively, were evaluated along with the parents in a randomized block design with a plot size of $2.0 \times 0.4 \text{ m}^2$ under rainfed (stress) conditions. The trials, DS01 and DS02 were evaluated with two and three replications respectively. Seeds were hand-dibbled into dry soil @ 80 kg/ha. NPK fertilizers were applied at the rate of 50:25:25 kg/ha. Insect and weed control measures were applied periodically, as required.

The trials, DS01 and DS02 received a total rainfall of 345 and 354 mm respectively, during crop growth. Table 1 gives the site, soil and drought stress characteristics of field trials. There was a dry spell for 30 days coinciding with flowering to grain filling in DS01. Perched soil water-table depth was measured at two-days interval using Peizometers (15×100 cm) made of polyvinyl carbonate in DS01. In DS02, there was no rainfall 40 days after emergence (DAE). Hence, one irrigation was given at 61 DAE at flowering stage to save the plants from desiccation. Since there was no rainfall after this irrigation, stress developed and continued till maturity. The drought stress was thus severe in DS02.

In FC01, the subset of 143 F_9 RI lines was evaluated along with the parents under irrigated conditions.

Twenty-six-day-old seedlings were transplanted with a spacing of 20×10 cm in plots of 1.2×0.2 m². The plots were surface-irrigated to field capacity at four days interval. NPK fertilizers were applied at the rate of 100:50:50 kg/ha.

In all the trials, observations for days to 50% flowering were recorded as the number of days from sowing to flowering in 50% of plants of each RI line. At maturity, plant height, number of tillers and spikelet number were recorded per plant. All the plants in the plot were harvested to record grain yield (kg ha⁻¹). Harvest index was estimated as the ratio of grain yield to the above-ground biomass. Drought score was recorded using leaf rolling at flowering stage during peak stress using 1–9 scale standardized for rice²⁶.

Genotyping

Leaf samples of RI lines and parents were collected from field-grown seedlings and freeze-dried. DNA was extracted from the leaf tissues using cetyl trimethyl ammonium borate buffer²⁷. The quantity and quality of DNA was assessed in 0.8% agarose gel and concentration was adjusted to 50 ng/µl by comparing DNA standards. Polymerase chain reaction (PCR) was performed in a volume of 15 µl in thermal cycler (DNA Engine[®], Peltier Thermal Cycler, BIO-RAD, USA or Eppendorf Master Cycler Gradient, Eppendorf, Germany). The reaction mixture contained 1 µM of each primer of (Sigma Aldrich, USA), 100 μ M deoxy nucleotide, 1× *Taq* buffer, 0.02 U Taq polymerase (Bangalore Genei, India) and 50 ng of template DNA. The PCR steps were as follows: initial denaturation for 5 min at 94°C, 36 cycles of amplification which included denaturation at 94°C for 1 min, annealing at 57°C for 1 min and extension at 72°C for 1 min and the final extension step at 72°C for 5 min. A set of 137 SSR markers in the QTL region (between RM585 and RM217) on chromosome 6 was selected from the rice physical map.

 Table 1. Site, soil and drought stress characteristics of field trials conducted in rainfed target populations of environment in Paramakudi (DS01 and DS02) and irrigated control in Coimbatore (FC01), India

| Particulars | DS01 (2011–2012) | DS02 (2012–2013) | FC01 (2012–2013) | |
|--|------------------|------------------|------------------|--|
| Elevation (m amsl) | 40 | 40 | 427 | |
| Latitude and longitude | 9°N, 70°E | 9°N, 70°E | 11°N, 77°E | |
| Soil texture | Clay loam | Clay loam | Clay | |
| Soil pH | 8.1 | 8.1 | 8.4 | |
| Characterization of stress | Moderate | Severe | Non-stress | |
| Total rainfall during crop period (mm) | 345 | 354 | _ | |
| Mean temperature (°C) | | | | |
| Maximum | 32.0 | 33.5 | 38.9 | |
| Minimum | 23.5 | 23.0 | 26.0 | |
| Average relative humidity (%) | 83.5 | 84.0 | 86.0 | |

DS01, Drought stress 2011–12; DS02, Drought stress 2012–13; FC01, Flooded control 2013.

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Statistical analysis

Analysis of variance for each trait in different trials was done as mixed models using PROC MIXED procedure of SAS V.9.3 (SAS Institute Inc.)²⁸. Combined analysis of the two dry season trials (DS01 and DS02) was done using PROC MIXED procedure of SAS, where the RI lines were kept as fixed and other variables were assigned as random. Broad sense heritability of different traits was estimated within a year for each treatment. The variance components for calculating broad sense heritability for each trait were calculated using SAS program PROC VARCOMP with REML method. PROC CORR and PROC SUMM were used for correlation and summary statistics calculations for all the trials in SAS.

Linkage map and QTL analysis

Linkage maps were constructed from the genotype data using MAPMAKER/EXP 3.0 software²⁹. The genetic distance was estimated using the Kosambi map function of the software³⁰. Markers were placed onto linkage groups using the group, order and ripple command for checking the critical region with a minimum logarithm of the odds (LOD) score of 3.0 and a maximum recombination fraction of 0.5 and the ordered linkage groups were drawn using MapChart³¹. QTL analysis for each trial was carried out with the line mean of phenotypic traits. Putative QTLs were detected using composite interval mapping (CIM) function of QTL Cartographer 2.5 (ref. 32). The CIM threshold was based on the results of 1000 permutations at 5% significance level³³. The additive effect and phenotypic variance explained by each QTL (R^2) were estimated at the maximum LOD score. QTLs with LOD score of 3 and above are reported.

Fine-mapping

The grain-yield QTL under drought on chromosome 6 between RM585 and RM217, previously identified from our laboratory³⁴ and also confirmed in the present study was chosen for fine-mapping. A total of 137 single sequence repeat (SSR) markers in the QTL region (between RM585 and RM217) on chromosome 6 were selected from the Rice Gene Thresher³⁵, and used in surveying polymorphism between the parents, IR62266 and Norungan. Twenty-seven polymorphic markers (see Supplementary material, Table S1 online) were used in genotyping the 143 RI lines to fine-map the QTL. Graphical genotyping software GGT 2 was used to draw genotypic graphs in RI lines³⁶.

Comparative genomics

Comparative genomics analysis was performed to identify the syntenic regions in maize, sorghum, pearl millet and barley using the genomics database (www. gramene.org), for the QTL region fine-mapped in this study. Syntenic regions identified were checked for the presence of grain-yield QTLs under stress. QTL information reported for grain yield was collected from the published literature and the Gramene database (www. gramene.org) and compared with the fine-mapped region using the comparative maps available in database.

Results and discussion

Variation in phenology and plant production traits

In this study, the landrace, Norungan, was used as a donor for grain yield under drought to identify consistent large-effect QTLs for plant production traits under drought stress in TPE. In DS01, the soil water table measured using peizometers declined below 80 cm starting from 61 DAE and reduced further to 95 cm (data not shown), coinciding with flowering to grain filling stages. In DS02, there was no rainfall beginning 40 DAE and remained dry till maturity, except for one irrigation given at 61 DAE to save the crop from total desiccation. Considerable variation was observed among the RI lines for phenology and plant production traits under drought stress and non-stress conditions.

The expression of genetic variability for droughttolerance traits requires a mapping population to be exposed to drought stress predominant in TPE. High-level water stress is desirable to remove the spillover effect of yield potential and identify the lines which are clearly drought-resistant^{12,20,37}. Table 2 provides the traits mean, range and broad sense heritability. Grain yield ranged from 31 to 2750 and 66 to 2889 kg/ha with mean of 940 and 644 kg/ha in DS01 and DS02 respectively. About 65-85% reduction in yield is considered severe drought¹³. In the present study, the reduction in mean grain yield was 80.4% and 86.2% in DS01 and DS02 respectively (Table 2). Such severe stress is useful in discriminating the superior lines under drought stress²⁰. The mean grain yield in FC01 (non-stress trial) was 4796 kg/ha. Heritability of yield was 0.70, 0.36 and 0.97 in the trails DS01, DS02 and FC01 respectively. Moderate to high heritability of grain yield under drought stress in DS01 and DS02 indicates the suitability of grain yield under drought as a selection criterion^{10,13}. Mean days to 50% flowering across the RI lines were 80, 97 and 90 in DS01, DS02 and FC01 respectively. Mean spikelet fertility across the RI lines was 57%, 72% and 84% in DS01, DS02 and FC01 respectively. Since the life irrigation given at flowering stage for the DS02 trial, the spikelet fertility was higher in this trial compared to DS01, but the drought that occurred before and after flowering affected the total grain yield in DS02; thus a lower mean grain yield was observed in DS02. [D1]The mean drought score was 3.8

| Trait | Trial | Norungan | IR62266 | Trial mean | Range | SD | H^2 |
|------------------------|-------|----------|---------|------------|-------------|-------|-------|
| Days to 50% flowering | DS01 | 76 | 80 | 80 | 58–95 | 10.45 | 0.73 |
| | DS02 | 105 | 108 | 97 | 61-108 | 8.41 | 0.18 |
| | FC01 | 78 | 102 | 90 | 78-122 | 8.64 | NA |
| Drought score | DS01 | 3 | 2 | 3.76 | 1–7 | 1.84 | 0.69 |
| | DS02 | 4 | 5 | 4.00 | 3–9 | 0.76 | 0.31 |
| | FC01 | _ | _ | _ | - | _ | - |
| Plant height (cm) | DS01 | 88 | 66 | 87 | 19–149 | 36.06 | 0.72 |
| | DS02 | 56 | 53 | 52 | 22-77 | 12.65 | 0.60 |
| | FC01 | 140 | 91 | 100 | 59-142 | 20.40 | 0.68 |
| No. of tillers | DS01 | 7 | 5 | 8 | 1–19 | 1.23 | 0.43 |
| | DS02 | 5 | 3 | 4 | 2-11 | 1.12 | 0.23 |
| | FC01 | 10 | 6 | 11 | 5-21 | 2.38 | 0.79 |
| No. productive tillers | DS01 | 7 | 7 | 8 | 1-17 | 3.00 | 0.34 |
| | DS02 | 4 | 3 | 4 | 1–9 | 1.33 | 0.20 |
| | FC01 | 10 | 6 | 11 | 3–24 | 2.49 | NA |
| No. of grains/panicle | DS01 | 44 | 46 | 45 | 3-100 | 19.40 | 0.35 |
| | DS02 | 74 | 83 | 26 | 9–106 | 9.75 | 0.07 |
| | FC01 | 62 | 127 | 82 | 17-160 | 31.17 | NA |
| No. of chaffs/panicle | DS01 | 27 | 32 | 32 | 4-71 | 13 | 0.42 |
| | DS02 | 8 | 15 | 9 | 2-24 | 3.69 | 0.10 |
| | FC01 | 8 | 5 | 19 | 1-64 | 13.02 | NA |
| Spikelet fertility (%) | DS01 | 62 | 66 | 57 | 26-83 | 13 | 0.40 |
| | DS02 | 90 | 90 | 72 | 31-90 | 10.52 | 0.21 |
| | FC01 | 89 | 96 | 84 | 51-99 | 11.50 | NA |
| Grain yield (kg/ha) | DS01 | 2000 | 1042 | 940 | 31-2750 | 734 | 0.70 |
| | DS02 | 1074 | 1007 | 644 | 66-2889 | 360 | 0.36 |
| | FC01 | 3750 | 5722 | 4796 | 813-13847 | 2529 | 0.97 |
| Straw yield (kg/ha) | DS01 | 6104 | 9400 | 7826 | 1219-64063 | 444 | 0.40 |
| | DS02 | 1127 | 1735 | 3491 | 685-9788 | 1543 | 0.27 |
| | FC01 | 17111 | 3014 | 9606 | 2333-22250 | 4071 | 0.99 |
| Total biomass (kg/ha) | DS01 | 8104 | 10442 | 8643 | 625-65313 | 5687 | 0.38 |
| | DS02 | 2296 | 3188 | 4112 | 981-27675 | 1730 | 0.81 |
| | FC01 | 20861 | 8736 | 13608 | 3403-33569 | 5698 | 0.96 |
| HI | DS01 | 0.25 | 0.15 | 0.12 | 0.01-0.30 | 0.08 | 0.55 |
| | DS02 | 0.40 | 0.32 | 0.17 | 0.01-0.43 | 0.08 | 0.99 |
| | FC01 | 0.18 | 0.66 | 0.36 | 0.04 - 1.00 | 0.19 | 0.97 |

 Table 2.
 Trait mean, range and broad sense heritability for phenology and plant production traits under drought stress (DS01 and DS02) and non-stress (FC01) conditions for IR62266 × Norungan RI lines and parents

HI, Harvest index; H^2 , Broad sense heritability; DS01, Drought stress 2011–12; DS02, Drought stress 2012–13; FC01, Flooded control 2013; NA, Not available.

and 4.0 in DS01 and DS02 respectively (Table 2). The drought score observed here shows the severity of stress in these trials.

QTL analysis

A total of 51 QTLs were identified in DS01, DS02 and FC01 among which 39 QTLs were observed in rainfed environment and 12 in non-stress environments (Table 3). Grain yield QTLs were identified on chromosomes 1, 6 and 10. Consistent large-effect QTL (qDTY6.1) was observed for grain yield under stress conditions in TPE on chromosome 6 near marker RM2434 with phenotypic variation of 31.3%, 37.9% and 3.6% in DS01, DS02 and FC01 respectively, in these RI lines. The same region was reported to be linked to grain yield³⁸, biomass, number of panicles¹² and thousand grain weight³⁹ in rice. QTL for

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seedling drought tolerance was reported near RM217 region, and the same region harbours genes expressed early under drought stress⁴⁰ and Khowaja *et al.*⁴¹ also reported QTLs for drought avoidance in rice in this region. A metaQTL for yield was reported near this region between RM204 and RM136 on chromosome 6 in rice⁴².

RM9 on chromosome 1 harboured major QTLs for grain yield, biomass and straw yield with a phenotypic variation of 38.9%, 37.4% and 34.2% respectively (Table 3). Association of RM9 with yield and yield components under stress has been reported earlier in rice⁴³⁻⁴⁶. RM5746 on chromosome 12 was linked to days to 50% flowering with a phenotypic variation of 69.77% in DS02. This region was also reported for root volume under drought stress⁴⁷. RM2434 on chromosome 6 and RM5746 on chromosome 12 were linked to drought score showing a phenotypic variation of 72% and 66% in DS01

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| | | Chromosome | Marker | Peak | Logarithm of | Additive | |
|---------------------------|-------|------------|---------------------|---------------|----------------|----------|-------|
| Trait | Trial | no. | closest to the peak | position (cM) | the odds (LOD) | effect | R^2 |
| Days to 50% flowering | DS01 | 12 | RM5746 | 44.01 | 11.46 | 1.53 | 6.5 |
| | | 6 | RM2434 | 72.83 | 7.34 | -3.42 | 8.6 |
| | DS02 | 12 | RM5746 | 50.01 | 7.26 | -10.30 | 69.7 |
| | | 6 | RM402 | 41.00 | 8.94 | -3.06 | 5.5 |
| | | 6 | RM2434 | 72.83 | 7.34 | -3.42 | 8.6 |
| | FC01 | 1 | RM8051 | 17.22 | 5.56 | -5.41 | 11.9 |
| | | 12 | RM5746 | 60.01 | 3.14 | 1.94 | 2.8 |
| Drought score | DS01 | 6 | RM2434 | 86.83 | 4.40 | 1.75 | 1.7 |
| | | 12 | RM5568 | 48.01 | 3.78 | -0.16 | 0.5 |
| | DS02 | 6 | RM2434 | 84.83 | 3.40 | 1.12 | 72.1 |
| | | 12 | RM5746 | 60.01 | 4.00 | 1.21 | 65.9 |
| Plant height (cm) | DS01 | 1 | RM8051 | 9.22 | 4.68 | 10.31 | 1.6 |
| | | 4 | RM6314 | 112.49 | 9.05 | -4.37 | 2.0 |
| | | 6 | RM402 | 35.91 | 6.96 | -19.06 | 8.7 |
| | | 6 | RM7561 | 26.01 | 7.46 | -10.75 | 4.2 |
| | | 10 | RM5471 | 7.8 | 7.19 | -19.27 | 11.6 |
| | | 10 | RM5095 | 2.01 | 8.55 | -20.06 | 9.2 |
| | FC01 | 6 | RM6773 | 86.80 | 7.06 | 5.73 | 3.8 |
| No. of tillers | FC01 | 2 | RM3512 | 17.76 | 3.60 | 0.59 | 5.7 |
| | | 2 | RM4472 | 17.62 | 3.70 | 0.58 | 5.7 |
| | | 2 | RM575 | 19.80 | 5.32 | 0.59 | 5.5 |
| No. of productive tillers | FC01 | 2 | RM575 | 21.80 | 4.15 | 0.58 | 4.7 |
| No. of chaffs | FC01 | 6 | RM6773 | 94.80 | 3.40 | 5.13 | 7.8 |
| Spikelet fertility (%) | DS02 | 2 | RM6942 | 76.53 | 3.36 | -2.14 | 3.7 |
| | FC01 | 6 | RM6773 | 94.80 | 9.48 | -3.79 | 5.3 |
| Single plant yield (g) | DS01 | 6 | RM6773 | 118.8 | 3.44 | -0.73 | 6.4 |
| | | 10 | RM5471 | 9.8 | 6.90 | -2.51 | 26.7 |
| | DS02 | 1 | RM9 | 15.22 | 4.43 | -3.50 | 23.1 |
| | | 4 | RM6679 | 72.16 | 3.90 | 1.54 | 13.5 |
| | | 4 | RM6314 | 86.49 | 4.03 | 1.60 | 14.1 |
| | FC01 | 6 | RM6773 | 88.88 | 3.81 | 5.24 | 15.1 |
| Grain yield (kg/ha) | DS01 | 1 | RM8051 | 15.22 | 3.52 | 252.69 | 3.2 |
| | | 6 | RM2434 | 86.83 | 3.90 | 629.2 | 31.3 |
| | | 6 | RM6773 | 91.45 | 3.06 | 0.04 | 2.5 |
| | | 10 | RM5471 | 11.8 | 6.82 | -584.05 | 25.1 |
| | DS02 | 6 | RM402 | 41.00 | 7.14 | 250.22 | 16.7 |
| | | 6 | RM2434 | 78.83 | 6.90 | 424.9 | 37.9 |
| | | 6 | RM6773 | 93.16 | 14.10 | 72.25 | 3.4 |
| | | 1 | RM9 | 9.22 | 13.45 | -54.60 | 38.9 |
| | FC01 | 6 | RM6773 | 84.80 | 5.10 | 638.07 | 3.6 |
| Straw yield (kg/ha) | DS01 | 3 | RM517 | 8.01 | 36.19 | 0.06 | 2.6 |
| | DS02 | 1 | RM9 | 13.22 | 11.10 | 0.12 | 34.2 |
| | | 11 | RM202 | 0.01 | 5.35 | 0.09 | 15.7 |
| Total biomass (kg/ha) | DS01 | 6 | RM6194 | 128.72 | 28.32 | -577.85 | 2.3 |
| | DS02 | 1 | RM9 | 11.22 | 15.68 | 0.07 | 37.4 |
| | | 6 | RM2434 | 76.83 | 11.85 | 583.00 | 5.7 |
| | | 11 | RM202 | 0.01 | 5.63 | 0.13 | 16.4 |
| | FC01 | 6 | RM6194 | 130.72 | 4.03 | 0.06 | 6.00 |
| HI | DS02 | 1 | RM9 | 13.22 | 3.45 | -0.04 | 5.6 |
| | | 12 | RM5746 | 82.01 | 3.10 | 0.03 | 10.1 |
| | FC01 | 6 | RM2434 | 72.91 | 9.86 | 0.12 | 16.6 |

Table 3. Quantitative trait loci (QTLs) for phenology and plant production traits mapped in IR62266/Norungan recombinant inbred lines under
drought stress (DS01 and DS02) non-stress (FC01) conditions

Peak position, Marker or position with the highest LOD value in the QTL curve; HI, Harvest index; R^2 , Broad sense heritability; DS01, Drought stress 2011–12; DS02, Drought stress 2012–13; FC01, Flooded control 2013.

and DS02 respectively. RM5471 on chromosome 10 was linked to grain yield under stress with the phenotypic variation of 25% in DS01. QTLs for osmotic adjustment capacity⁴⁸ and grain weight⁴¹ were reported near RM5471 on chromosome 10 in rice.

Fine mapping of qDTY6.1

The consistent large-effect QTL for yield under drought stress in TPE mapped earlier in these RI lines between the flanking markers RM585 and RM217 on chromosome

6 (qDTY6.1) with an interval of ~3.0 Mb was chosen for fine-mapping³⁴. The candidate genomic region was finemapped to the smallest fragment of 94.04 kb between RM2434 and RM6773. To define the candidate genomic region for qDTY6.1, 137 SSRs were selected from the interval between RM585 and RM217 based on the physical map position in Rice Gene Thresher³⁵. Among these, 27 markers which showed polymorphism between the parents (IR62266 and Norungan) were used in genotyping the 143 RI lines. CIM analysis identified the region flanked by RM2434 and RM6773 on chromosome 6 to be consistently linked to grain yield under stress conditions (Figure 1). qDTY6.1 had its peak at 86.8 cM, close to the marker RM2434. The consistent large-effect qDTY6.1 was fine-mapped between RM2434 (4310.93 kb) and RM6773 (4404.98 kb) with an interval of 94.04 kb (Figure 2). The presence of genomic segment between RM2434 and RM6773 (qDTY6.1) shows a positive effect on grain yield under stress and non-stress conditions.



Figure 1. Consistent quantitative trait locus (QTL) peaks on finemapped region between RM2434 and RM6773 in stress (DS01 and DS02) and non-stress (FC01) trials: the curve between the mentioned markers is the LOD score for the grain yield QTL, qDTY6.1. R^2 indicates the phenotypic variance explained in percentage. AE, Additive effect. *a*, Stress condition during the year 2011–12 (DS01). *b*, Stress condition during the year 2012–13 (DS02). *c*, Non-stress condition during the year 2013–14 (FC01).

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The positive alleles were contributed by the landrace, Norungan. Thus the region qDTY6.1 is a potential candidate for improving rainfed rice yields through deployment in MAB and/or map-based cloning of underlying genes.

Rice Gene Thresher³⁵ was used to mine this region with stress gene catalogue and identified 21 genes that were upregulated under drought stress condition. Plastocyanin, vacuolar proton pyrophosphatase, acyl-CoAbinding protein and phosphate-induced protein-1 are Dayhoff's curated stress protein family present in the qDTY6.1 region. These proteins are related to photosynthesis^{49,50}, development of larger root system with increased auxin transport or higher auxin concentration in the root system with increased yield under dryland condition. ACPB2 was induced by abscisic acid (ABA) and drought. ABA-mediated reactive oxygen species (ROS) production in guard cells promotes stomatal closure, reduces water loss and enhances drought tolerance. Overexpression of ACBP2 improves drought tolerance and the mutant acpb2 is more sensitive to drought stress⁵¹.

Comparative genomics of qDTY6.1

The evolutionary relationship among cereals is a known fact and the syntenic relationship can be used to find the homologous regions, which are useful in identifying their significance in stress adaptation in cereals. We compared the fine-mapped qDTY6.1 region for synteny among other cereals. The qDTY6.1 region was syntenic with maize (see Supplementary material, Figure S1 a online), pearl millet, sorghum (see Supplementary material, Figure S1 b online) and barley. This region was also found in



Figure 2. Location of qDTY6.1 on chromosome 6. (*a*) Linkage map constructed in a previous study³⁴ and distance in physical map position. *b*, Fine-mapped region (qDTY6.1) between the markers RM2434 and RM6773 and its narrowed distance in the physical map.

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maize on chromosome 9 near the marker umc109 in bin-9.02, sorghum on linkage group F, HHU37a, HHU37b and HHU37c, pearl millet on chromosome 2, Xpsr490(Ss1) and barley on chromosome 7H near marker ABC255 in bin-6. The syntenic regions in maize⁵², pearl millet⁵³, sorghum⁵⁴ and barley^{55,56} were reported for grain yield under drought stress. In sorghum, the syntenic region was linked to photosynthesis-related vacuolar H+translocating pyrophosphatase (see Supplementary material, Figure S1 *b* online).

Conclusion

To sum up, a consistent large-effect QTL (qDTY6.1) for yield under drought predominant in TPE was further confirmed and fine-mapped for application in MAB using rice lines derived from a local land race, Norungan. This region is also syntenic among cereals, indicating that drought adaptation mechanisms underlying this QTL have been conserved across cereals. qDTY6.1 is a potential candidate for improving grain yield under water stress in rice and other cereals as well. Further cloning of genes in this region may be useful in improving the production of rice and other cereal under water-limited environments.

- Food and Agriculture Organization, 2012; http://www.fao.org/ economic/est/publications/rice-publications/rice-market-monitorrmm/en/
- Food and Agriculture Organization, 2013; http://www.fao.org/ economic/est/publications/rice-publications/rice-market-monitorrmm/en/
- Hanson, A. D., Peacock, W. J., Evans, L. T., Arntzen, C. J. and Khush, G. S., Drought resistance in rice. *Nature*, 1990, 345, 26–27.
- Pandey, S. and Bhandari, H., Introduction. In Economic Costs of Drought and Rice Farmers' Coping Mechanisms. A Cross-Country Comparative Analysis (eds Pandey, S., Bhandary, H. and Hardy, B), International Rice Research Institute/World Scientific, Singapore, 2007, pp. 1–9.
- Kumar, A., Dixit, S., Ram, T., Yadaw, R. B., Mishra, K. K. and Mandal, N. P., Breeding high-yielding drought-tolerant rice: genetic variations and conventional and molecular approaches. *J. Exp. Bot.*, 2014, 65, 6265–6278.
- Jongdee, B., Fukai, S. and Cooper, M., Genotypic variation for grain yield of rice under water-deficit conditions. In Proceedings of 9th Australian Agronomy Conference (eds Michalk, D. and Pratley, J.), Wagga Wagga, NSW, Australia, 1998, pp. 403–406.
- Salunkhe, A. S. *et al.*, Fine mapping QTL for drought resistance traits in rice (*Oryza sativa* L.) using bulk segregant analysis. *Mol. Biotechnol.*, 2011, 49, 90–95.
- Dixit, S., Singh, A. and Kumar, A., Rice breeding for high grain yield under drought: a strategic solution to a complex problem. *Int. J. Agron.*, 2014, 863683, 13; doi:10.1155/2014/863683.
- Kumar, A., Bernier, J., Verulkar, S., Lafitte, H. R. and Atlin, G. N., Breeding for drought tolerance: direct selection for yield, response to selection and use of drought-tolerant donors in upland and lowland-adapted populations. *Field Crops Res.*, 2008, **107**, 221–231.
- Venuprasad, R., Lafitte, H. R. and Atlin, G. N., Response to direct selection for grain yield under drought stress in rice. *Crop* Sci., 2007, 47, 285–293.

- 11. Xu, Y. B., *Molecular Breeding*, CAB International, Wallingford UK, 2010.
- Bernier, J., Kumar, A., Ramaiah, V., Spaner, D. and Atlin, G., A large-effect QTL for grain yield under reproductive-stage drought stress in upland rice. *Crop Sci.*, 2007, 47, 507–518.
- Kumar, R., Venuprasad, R. and Atlin, G. N., Genetic analysis of rainfed lowland rice drought tolerance under naturally-occurring stress in eastern India: heritability and QTL effects. *Field Crops Res.*, 2007, **103**, 42–52.
- Venuprasad, R., Bool, M. E., Dalid, C. O., Bernier, J., Kumar, A. and Atlin, G. N., Genetic loci responding to two cycles of divergent selection for grain yield under drought stress in a rice breeding population. *Euphytica*, 2009, 167, 261–269.
- Vikram, P., Swamy, B. P. M., Dixit. S., Ahmed, H. U., Sta Cruz, M. T., Singh, A. K. and Kumar, A., qDTY_{1.1}, a major QTL for rice grain yield under reproductive-stage drought stress with a consistent effect in multiple elite genetic backgrounds. *BMC Genet.*, 2011, **12**(1), 89.
- 16. Mishra, K. K. *et al.*, qDTY_{12.1}: a locus with a consistent effect on grain yield under drought in rice. *BMC Genet.*, 2013, **14**, 12.
- Courtois, B. *et al.*, Locating QTLs controlling constitutive root traits in the rice population IAC165xCo39. *Euphytica*, 2003, 134, 335–345.
- Salekdeh, G. H., Siopongco, J., Wade, L. J., Ghareyazie, B. and Bennett, J., Proteomic analysis of rice leaves during drought stress and recovery. *Proteomics*, 2002, 2, 1131–1145.
- Dixit, S. *et al.*, Fine mapping of QTLs for rice grain yield under drought reveals sub-QTLs conferring a response to variable drought severities. *Theor. Appl. Genet.*, 2012, **125**, 155–169.
- Lafitte, H. R. *et al.*, Improvement of rice drought tolerance through backcross breeding: evaluation of donors and selection in drought nurseries. *Field Crops Res.*, 2006, **97**, 77–96.
- Atlin, G. N., Lafitte, H. R., Tao, D., Laza, M., Amante, M. and Courtois, B., Developing rice cultivars for high-fertility upland systems in the Asian tropics. *Field Crops Res.*, 2006, 97, 43–52.
- Tanksley, S. D. and McCouch, S. R., Seed banks and molecular maps: unblocking genetic potential from the wild. *Science*, 1997, 277, 1063–1066.
- Babu, R. C. *et al.*, Variation in root penetration ability, osmotic adjustment and dehydration tolerance among accessions of rice adapted to rainfed lowland and upland ecosystems. *Plant Breed.*, 2001, **120**, 233–238.
- Babu, R. C., Pathan, M. S., Blum, A. and Nguyen, H. T., Comparison of measurement methods of osmotic adjustment in rice cultivars. *Crop Sci.*, 1999, **39**, 150–158.
- Siopongco, J. D. L. C., Sekiya, K., Yamauchi, A., Egdane, J., Ismail, A. M. and Wade, L. J., Stomatal responses in rainfed low-land rice to partial soil drying; comparison of two lines. *Plant Prod. Sci.*, 2009, **12**, 17–28.
- IRRI, International network for genetic evaluation of rice: standard evaluation system for rice. International Rice Research Institute, Los Banos, the Philippines, 1996.
- Gawel, N. J. and Jarret, R. L., A modified CTAB DNA extraction procedure for *Musa* and *Ipomoea* plant. *Mol. Biol. Rep.*, 1991, 9, 262–266.
- SAS Institute Inc, SAS® 9.3. SAS Institute Inc, Cary, NC, USA, 2012.
- Lander, E. S., Green, P., Abrahamson, J., Barlow, A., Daly, M. J., Lincoln, S. E. and Newberg, L., MAPMAKER: an interactive computer package for constructing primary genetic linkage maps of experimental and natural populations. *Genomics*, 2009, 1, 174– 181.
- Kosambi, D. D., The estimation of map distances from recombination values. *Ann. Eugenet.*, 1944, **12**(1), 172–175.
- 31. Voorrips, R. E., MapChart: software for the graphical presentation of linkage maps and QTLs. *J. Hered.*, 2002, **93**, 77–78.

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- Wang, S., Basten, J. and Zeng, Z. B., Windows QTL Cartographer 2.5. North Carolina State University, Raleigh, NC, 2013.
- 33. Churchill, G. A. and Doerge, R. W., Empirical threshold values for quantitative trait mapping. *Genetics*, 1994, **138**, 963–971.
- Suji, K. K. *et al.*, Mapping QTLs for plant phenology and production traits using indica rice (*Oryza sativa* L.) lines adapted to rainfed environment. *Mol. Biotechnol.*, 2012, **52**, 151–160.
- Thongjuea, S., Ruanjaichon, V., Bruskiewich, R. and Vanavichit, A., RiceGeneThresher: a web-based application for mining genes underlying QTL in rice genome. *Nucleic Acids Res.*, 2009, 37, D996–D1000.
- 36. Berloo, R. V., GGT 2.0: versatile software for visualization and analysis of genetic data. *J. Hered.*, 2008, **99**, 232–236.
- Babu, R. C. *et al.*, Genetic analysis of drought resistance in rice by molecular markers: association between secondary traits and field performance. *Crop Sci.*, 2003, **43**, 1457–1469.
- Cho, Y. G. *et al.*, Identification of quantitative trait loci in rice for yield, yield components and agronomic traits across years and locations. *Crop Sci.*, 2007, 47, 2403–2417.
- 39. Zhuang, J. Y., Fan, Y. Y., Wu, J. L., Xia, Y. W. and Zheng, K. L., Comparison of the detection of QTL for yield traits in different generations of a rice cross using two mapping approaches. *Yi Chuan Xue Bao*, 2001, **28**, 458–464.
- 40. Zhou, S. X., Tian, F., Zhu, Z. F., Fu, Y. C., Wang, X. K. and Sun, C. Q., Identification of quantitative trait loci controlling drought tolerance at seedling stage in Chinese Dongxiang common wild rice (*Oryza rufipogon* G.). Acta Genet. Sin., 2006, 33, 551–558.
- Khowaja, F. S., Norton, G. J., Courtois, B. and Price, A. H., Improved resolution in the position of drought-related QTLs in a single mapping population of rice by meta-analysis. *BMC Genomics*, 2009, 10, 276–290.
- 42. Swamy, B. P. M. and Sarla, N., Meta-analysis of yield QTLs derived from inter-specific crosses of rice reveals consensus regions and candidate genes. *Plant Mol. Biol. Rep.*, 2010, **29**, 663–680.
- Cho, Y. C. *et al.*, QTLs analysis of yield and its related traits in wild rice relative *Oryza rufipogon*. *Treat Crop Res.*, 2003, 4, 19– 29.
- 44. Brondani, C., Rangel, N., Brondani, V. and Ferreira, E., QTL mapping and introgression of yield-related traits from Oryza glumaepatula to cultivated rice (Oryza sativa) using microsatellite markers. Theor. Appl. Genet., 2002, 104, 1192–1203.
- 45. Marri, P. R., Sarla, N., Reddy, L. V. and Siddiq, E., Identification and mapping of yield and yield related QTLs from an Indian accession of *Oryza rufipogon*. *BMC Genet.*, 2005, **6**, 33.
- 46. Moncada, P., Martinez, C. P., Borrero, J., Chatel, M., Gauch Jr, H., Guimaraes, E., Tohme, J. and McCouch, S. R., Quantitative

trait loci for yield and yield components in an *Oryza sativa* × *Oryza rufipogon* BC2F2 population evaluated in an upland environment. *Theor. Appl. Genet.*, 2001, **102**, 41–52.

- 47. Qu, Y., Mu, P., Zhang, H., Chen, C.Y., Gao, Y., Tian, Y., Wen, F. and Li, Z., Mapping QTLs of root morphological traits at different growth stages in rice. *Genetica*, 2008, **133**, 187–200.
- Robin, S. *et al.*, Mapping osmotic adjustment in an advanced back-cross inbred population of rice. *Theor. Appl. Genet.*, 2003, 107, 1288–1296.
- Li, W. *et al.*, Proteomics analysis of alfalfa response to heat stress. *PLoS ONE*, 2013, 8, e82725; doi:10.1371/journal.pone.0082725.
- Zhang, H., Shen, G., Kuppu, S., Gaxiola, R. and Payton, P., Creating drought- and salt-tolerant cotton by overexpressing a vacuolar pyrophosphatase gene. *Plant Signal Behav.*, 2011, 6, 861–863.
- Du, Z. Y., Chen, M. X., Chen, Q. F., Xiao, S. and Chye, M. L., Overexpression of *Arabidopsis* acyl-CoA-binding protein ACBP2 enhances drought tolerance. *Plant Cell Environ.*, 2013, 36, 300– 314.
- 52. Hao, Z. F., Li, X. H., Xie, C. X., Li, M. S., Zhang, D. G., Bai, L. and Zhang, S. H., Two consensus quantitative trait loci clusters controlling anthesis-silking interval, ear setting and grain yield might be related with drought tolerance in maize. *Ann. Appl. Biol.*, 2008, **153**, 73–83.
- 53. Yadav, R. S., Hash, C. T., Bidinger, F. R., Cavan, G. P. and Howarth, C. J., Quantitative trait loci associated with traits determining grain and stover yield in pearl millet under terminal drought-stress conditions. *Theor. Appl. Genet.*, 2002, **104**, 67–83.
- Wyrich, R. *et al.*, The molecular basis of C4 photosynthesis in sorghum: isolation, characterization and RFLP mapping of mesophyll- and bundle-sheath-specific cDNAs obtained by differential screening. *Plant Mol. Biol.*, 1998, **37**, 319–335.
- Forster, B. P. *et al.*, Genotype and phenotype associations with drought tolerance in barley tested in North Africa. *Ann. Appl. Biol.*, 2004, 144, 157–168.
- Von Korff, M., Grando, S., Del Greco, A., This, D., Baum, M. and Ceccarelli, S., Quantitative trait loci associated with adaptation to Mediterranean dryland conditions in barley. *Theor. Appl. Genet.*, 2008, **117**, 653–669.

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