Lipid links inflammation, immunity and insulin resistance to cause epidemic diabetes

Samir Bhattacharya* and Sandip Mukherjee

Cellular and Molecular Endocrinology Laboratory, Centre for Advanced Studies in Zoology, School of Life Science, Visva-Bharati, Santiniketan 731 235, India

Incidence of type 2 diabetes (T2D) is dramatically increasing in the past few decades and presently affecting more than 350 million people all over the globe. Oversupply of lipid is one of the major forces behind T2D. Excess of lipid decreases insulin sensitivity or activity that causes insulin resistance, a stage which occupies the centre of pathogenesis in T2D. Lipid induces adipose tissue inflammation that accompanies certain critical defects in adipocytes, a major cell in abdominal adipose tissue. These include increased population of hypertrophied adipocytes, decline in adipokines secretion, attenuation of adipogenesis, and increased lipotoxicity effecting greater deposition of fat which interferes with glucose uptake by insulin target cells. Inflammation of adipose tissue is further intensified due to the infiltration of macrophages, a member of the innate immune system, and their transformation from anti-inflammatory M2 to proinflammatory M1 phenotype. Hence, secretion of proinflammatory cytokines from both M1 macrophage and inflamed adipocytes is greatly elevated which adversely causes insulin resistance that leads to T2D. Association between lipid-induced inflammation and insulin resistance makes diabetes a critical disease.

Keywords: Adipose tissue inflammation, FetuinA, insulin resistance, macrophage infiltration, type 2 diabetes.

TYPE 2 DIABETES (T2D) apparently looks to be a simple disease where excess of lipid causes loss of insulin sensitivity which begins with insulin resistance. Initially, this defect in insulin activity could not be perceived by patient as the dysregulation of glycemic level is being prevented by the compensatory increase in insulin secretion from the pancreatic β -cell. Hence, it is hyperinsulinaemia that maintains the non-diabetic state with euglycaemia. This stage is termed as insulin resistance. But, a time comes when β -cell fails to bear this extra burden of hyperinsulinaemia and a decline in plasma insulin level results onset of T2D¹⁻³. In contrast, this stage is absent in type 1 diabetes where hyperglycaemia occurs due to the dearth of insulin secretion because of β -cell destruction caused by the autoimmune disorder¹. But this is not an epidemic disease as only 3-5% of diabetic population belong to this class whereas 95–97% are T2D, which is now posing a threat to global health⁴.

Hence, it is lipid or fat which induces loss of insulin sensitivity or inactivation of insulin. Several reports have demonstrated that removal of fat permits regaining of insulin sensitivity⁵⁻⁷. More precisely, long chain saturated free fatty acids (FFA) or non-esterified fatty acids (NEFA) have been shown to be the major player affecting the impairment of insulin activity. Last few years of research documented that excess of lipid is associated with chronic inflammation, particularly in the adipose tissue and this causes insulin resistance in different animal models^{4,8-9}. In human beings, obesity-induced chronic inflammation is the major factor for insulin resistance. Interestingly, increased macrophage infiltration in adipose tissue and their transformation to proinflammatory subtypes play a significant role in aggravating adipose tissue inflammation as observed with hyperlipidaemic diabetic patients¹⁰⁻¹³. In this review, we will primarily deal with the association between lipid-induced adipose tissue inflammation, participation of immune cells therein and insulin resistance that leads to the development of T2D.

Impairment of adipose tissue: the major regulator of insulin resistance

Abdominal white adipose tissue (WAT) has been recognized as an important site for storing energy obtained from diet. The predominant cell type in this tissue is adipocytes which store energy in the form of triglyceride (TG) as simple lipid droplets. During caloric need, TG is hydrolysed, resulting in free fatty acids, oxidation of which in the mitochondria, primarily in the skeletal muscle tissue produces energy in the form of adenosine triphosphate (ATP)^{14–18}. Adipocytes of WAT are now regarded very important cells, dysfunction of which leads to the extracellular accumulation of lipid, increase in circulatory lipid level and ectopic fat deposition (Figure 1). All these disrupt the balance in energy storage and expenditure, and impair energy homeostasis which is responsible for the decrease in insulin sensitivity that results in insulin resistance¹⁸. This is because adipocytes perform certain critical functions like uptake of fatty acids, storing them in the form of TG and converting them to free fatty acids (FFAs) through lipolysis during

^{*}For correspondence. (e-mail: bhattacharyasa@gmail.com)

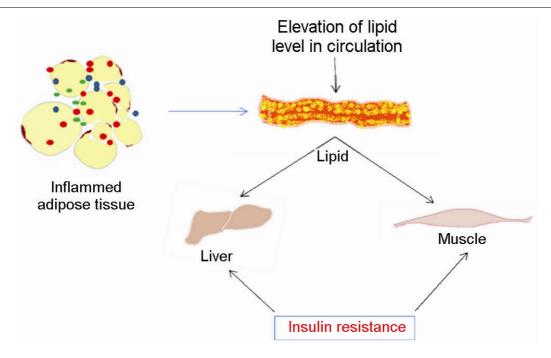


Figure 1. Fatty acid release from inflamed adipose tissue increases lipid level in circulation and effects fat deposition in liver and muscle causing insulin resistance.

energy demand. FFAs released into the circulation from adipocytes are taken up by the muscle, converted into TG and during energy crisis, TGs are hydrolysed to FFAs, and mobilized to mitochondria for β -oxidation to yield energy. It is intriguing to note that the capacity for storage of FFAs is far greater in adipocytes as compared to muscle cells but the ability for oxidation of FFAs in mitochondria is more efficiently operated in muscle cells than in adipocytes^{17,18}.

Interestingly, morphology of adipocytes varies according to their functional status; for example, populations of preadipocytes and small adipocytes are more in normal condition whereas large or hypertrophied and senescent adipocytes increase in obese diabetic subjects and animals^{19,20}. Hypertrophied and senescent adipocytes are considered to be aberrant in function and increase in their number is associated with insulin resistance and T2D²¹. In lean, non-diabetic mice or human beings, adipose tissue is largely occupied by small to middle sized adipocytes which are highly active and do not allow extracellular fat deposition^{22,23}. In contrast, during obese diabetic conditions, significant decline in pre- and small adipocytes is observed with a remarkable increase in hypertrophied adipocytes, which are defective cells, unable to uptake glucose and FFA appropriately and in addition, they extrude FFA affecting glucose and insulin intolerance that leads to T2D^{17,24}. Senescence in adipocytes, on the other hand, is the product of oxidative stress, sometimes occurring because of over-utilization of adipocytes. They are functionally inert, usually non-dividing but occupy the space in inflamed adipose tissue as observed in diabetic mice and human subjects^{21,25}

Increase in hypertrophied adipocyte population is one of the markers for the loss of insulin sensitivity which is related to impairment of PPAR γ (peroxisome proliferator activator receptor γ) expression and activity. PPAR γ is the key factor in improving lipid-induced insulin resistance $^{26-28}$. However, PPAR γ itself is not involved in insulin sensitization but it improves insulin action through the transcription of its target genes such as adiponectin, CD36 and aP2 (refs 27, 29, 30). Adiponectin, although secreted from adipocytes, regulates fatty acid mobilization and oxidation predominantly in muscle and therefore is an important adipokine for maintaining insulin sensitivity in muscle¹⁶. CD36 uptake fatty acid and also act as translocase; and aP2 is a fatty acid binding protein and regulates its intracellular mobilization^{31,32}. Negative regulation of PPARy and its target genes in adipocytes due to excess lipid therefore affects loss of insulin sensitivity causing insulin resistance. From this description, it would be evident that dysregulation of adipocyte function because of excess fat is a key factor in insulin resistance^{16,17}.

Fatty acid impairs insulin signalling that contributes to insulin resistance

Among some previously described information which demonstrated a direct association of FFA with insulin resistance, the most convincing evidence was available from the report of Santomauro *et al.*⁵. By performing some elegant experiments, they showed that lowering of FFA in T2D subjects significantly reduces insulin

resistance and strikingly improves oral glucose tolerance. Again, forcing FFA entry through infusion in rats and humans has been found to damage insulin sensitivity and produces insulin resistance^{23,33}. Blocking of FFA action or reduction of FFA level has been shown to improve insulin sensitivity. There are several studies which show that FFAs are responsible for insulin resistance which ultimately results $T2D^{2,4,34-37}$. FFA is now known to cause defects on insulin activity, which is contributed by several factors those adversely affect insulin responsive tissues such as muscles, liver and adipose tissues. One of such factors is insulin receptor (IR). Binding of insulin with IR initiates insulin signalling cascades which through downstream signalling components such as IRS1, PI3K, PDK1 and Akt activates Glut4 in adipocytes or skeletal muscle cells to uptake glucose from circulation into the cell. Failure of insulin stimulated signals therefore causes hyperglycaemia or increase in blood glucose level. There are both quantitative and qualitative defects of IR in T2D patients as the number of IR per adipocytes is significantly reduced in obese diabetic subjects, and decrease in IR gene expression has been detected in insulin-resistant patients³⁸⁻⁴². Reduced expression of IR in insulin responsive tissues of animals has also been shown to be affected due to FFAs, mainly by long chain saturated fatty acids. Decrease in IR protein and mRNA expression in hepatocytes and skeletal muscle cells occur due to FFAs such as palmitate which impairs insulin signalling^{41,43}. Palmitate, through the inhibition HMGA1 (high mobility group protein A1), an architectural transcription factor of IR gene, inhibits IR transcriptional activity that downregulates IR gene expression in muscle cells which compromises insulin activity⁴⁴.

Lipid-induced adipose tissue inflammation and insulin resistance

FFAs, especially long chain fatty acids, could trigger cellular proinflammatory pathways as demonstrated by several studies^{12,45–47}. FFAs mediate their proinflammatory effect primarily through the activation of toll-like receptors (TLRs)-dependent mechanisms which show that excess of lipid or lipotoxicity is associated with chronic inflammation of adipose tissue that causes decrease in insulin sensitivity^{12,45,48–50}. In the absence of TLR4, FFAmediated inflammatory signalling is suppressed in adipocytes⁴⁹. Moreover, insulin activity remains protected in TLR4 knockout mice against high fat diet⁵¹. FFA-induced activation of TLR4, a pattern recognition receptor, mediates inflammation through NF kB pathways 47,50,52-54 and it also stimulates JNK and IKK which suppresses insulin signalling through serine phosphorylation of IRS I (ref. 12). This indicates that FFA could act as a ligand of TLR4, and their binding causes insulin resistance in adipocytes. However, it has been shown that fatty acid has no direct association with TLR4 (ref. 55), hence it is critically important to know how FFA can activate TLR4 mediated inflammatory signal without binding to TLR4. It has been reported that excess FFA induces Fetuin A or α₂ Heremans–Schmid glycoprotein (AHSG) expression primarily in liver⁵⁶. In obesity-induced diabetic humans and mice, serum Fetuin A (FetA) level significantly increases as compared to non-diabetic subjects or mice^{57–61}. Interestingly, FetA directly binds to TLR4 with high affinity and it also binds to FFA very strongly. It has been found that FetA binds to FFA more avidly than albumin and then presents it to TLR4. FFA–FetA–TLR4 form a ternary complex which activates TLR4–NFkB pathways to produce proinflammatory cytokines from inflamed adipocytes which exacerbate insulin resistance⁶².

However, *FetA* gene is not only expressed in liver but also in adipocytes. Its expression is induced by fat and greater accumulation of lipid is correlated with the amount of *FetA* gene expression⁶³. FetA knockout mice are shown to be protected from high fat diet-induced insulin resistance⁶⁴ and in FetA knockdown mice, high fat diet fails to induce adipose tissue inflammation and loss of insulin sensitivity⁶². FetA's contribution to the development of metabolic disorders leading to insulin resistance and T2D including other complications has recently been dealt in detail by Trepanowski *et al.*⁶¹.

Adipocytes dictate impairment of insulin sensitivity in muscle and liver

Insulin resistance in muscle is largely controlled through adipocyte dysfunction because greater supply of lipid and attenuation of adiponectin secretion adversely affects insulin sensitivity of muscle tissue. Larger amounts of post-prandial glucose in blood after a meal is being deposited (~75%) in skeletal muscle tissues^{65–67}. Therefore, loss of insulin sensitivity in the muscles of human subject and mice produces significant effects on glucose homeostasis. Recent studies demonstrated that hyperlipidemic condition during obesity is correlated with enhanced expression of proinflammatory genes in the muscles of mice and human subjects^{13,68}. In addition, due to dysfunction of adipocytes, increased level of serum fatty acids markedly enhances FFA influx into the muscle of obese diabetic individuals which significantly attenuates fatty acid oxidation. This further contributes to the accumulation of fatty acid intermediates in muscle that activates JNK, IKK or nPKC signalling pathways causing insulin $resistance^{44,69-71} \\$

Contribution of adipose tissue for developing insulin resistance in muscle and liver involves adipokines. Adipocyte synthesizes 347 proteins, of which 263 are predicted to be secretory and adipokine in nature⁷². Major adipokines secreted from it (Figure 2) create a microenvironment through interconnected network which enable

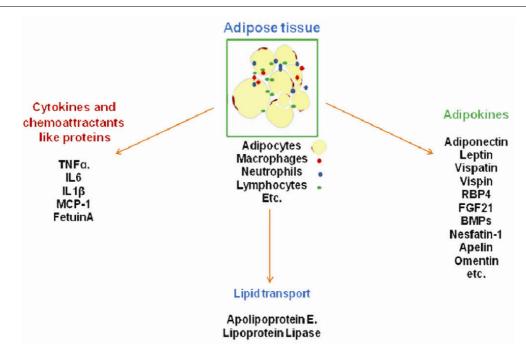


Figure 2. Adipose tissue secretes several proteins such as chemoattractants like proteins, apolipoproteins, adipokines, etc. crucial for maintaining energy homeostasis.

adipocytes to communicate with skeletal muscles and liver^{72,73}. During obese diabetic conditions, aberrant adipokine secretion from inflamed adipose tissue worsens the situation in muscle and liver. The most crucial reason behind such abnormality appears to be the excess efflux of FFA by hypertrophied adipocytes of obese diabetic mice and human subjects leading to considerable increase in circulatory FFA level and extracellular lipid accumulation. This causes insulin resistance in adipose tissue, muscle and liver^{3,6,7,34–37} and lowering of FFA increases insulin sensitivity in these tissues. A significant increase in muscle triglyceride (TG) concentration and hepatic steatosis are commonly occurring abnormalities observed during insulin resistance⁷⁴. FFA efflux from hypertrophied adipocytes when accumulated in liver and muscle may generate bioactive lipid products which can inhibit insulin signalling⁷⁵. In skeletal muscle cells, fatty acid derived diacylglycerides (DAGs) can augment protein kinase C (PKC) activity which inhibits serine phosphorylation of insulin receptor substrate (IRS) or FFA can directly stimulate PKC activation which leads to the downregulation of insulin receptor - both causing insulin resistance⁴⁴. From these studies, it is evident that abnormalities in adipocytes are transmitted to liver and muscle tissue via the excess lipid load which greatly contribute to the development of insulin resistance and T2D.

Another important dimension of excess FFA in skeletal muscle tissues is its effect on mitochondrial dysfunction which impairs insulin sensitivity. Decreased mitochondrial density and oxidative capacity of muscle have been detected in T2D patients^{76–78}. Even in healthy animals

and humans, high fat diet or lipid infusion markedly decreases oxidative phosphorylation and ATP synthesis in mitochondria $^{79-81}$. This indicates that FFAs can directly cause dysfunction of skeletal muscle mitochondria which is related to insulin resistance. Several reports also suggest that intramuscular increase of FFAs and triglyceride levels may produce defects in mitochondrial β -oxidation which is associated with insulin resistance $^{82-84}$.

Macrophage participation in adipose tissue inflammation: a critical stage in insulin resistance

It is now increasingly recognized that the immune system and metabolic disorder leading to insulin resistance and T2D are highly integrated. High energy diet and accumulation of excess lipid are affecting lipotoxicity, triggers a number of metabolic pathways responsible for the development of insulin resistance. One of the critically important pathogenic inputs in insulin resistance is macrophage infiltration into inflamed adipose tissue which produces proinflammatory cytokines that further intensify inflammation and worsen insulin sensitivity 9-12,45. It is interesting to note that inflammation of adipose tissue due to excess lipid attracts macrophage. Migration of macrophage has been reported to be mediated by Monocyte Chemoattractant Protein1 (MCP1)⁴⁶, which is found to be highly expressed in adipose tissue and secreted in HFD mice^{13,85}. In addition, MCP1 and MCP1 receptor (CCR2) have shown to be important in the recruitment of macrophages into the adipose tissue.

The most remarkable dimension of adipose tissue inflammation is related to resident adipose tissue macrophage (ATM). In obesity-induced inflammation, accumulation of macrophage in the adipose tissue is a characteristic feature of chronic inflammation which causes insulin resistance and $T2D^{10-12}$. Why inflamed adipose tissue is an attractive site for macrophage infiltration is yet unclear. No doubt MCP1 is a chemoattractant for macrophage migration but CCR2 deficient mice have been shown to be protected from HFD-induced insulin resistance and macrophage accumulation 85,86. Moreover, a few reports indicate that MCP1 is not sufficient for the amount of macrophage that infiltrates during adipose tissue inflammation⁸⁷. Hence there appears to be other chemoattractant(s) necessary for such purpose. Most conspicuous lacunae in this field is in the understanding of the transformation of M2 or classically activated macrophage (CAM) to M1 or alternative activated macrophage (AAM) which contributes to severe inflammation that significantly reduce insulin sensitivity¹³. It has been recently reported that FetA released from adipocytes is also acting as chemoattractant for macrophage infiltration to the adipose tissue and most interestingly, FetA also contributes to the polarization of M2 to M1 phenotypes⁶³. Till date, no other factor has been detected to influence M2 to M1 transformation. The microenvironment thus created by inflamed adipocytes and pro-inflammatory M1 macrophage cohabitation could produce aberrant metabolic functions that result in an unusual increase in the release of proinflammatory cytokines such as TNF α and IL6 (refs 12, 13). In this complex inflammatory condition, migration and accumulation of macrophages in the adipose tissue are reported to be the principal source of inflammatory mediators through the expression and release of TNF $\alpha^{10,11,45}$. Excess secretion of TNF α from both adipose tissue and macrophages is a strong negative regulator of insulin sensitivity, potent enough to cause insulin resistance. Moreover, TNF α augments lipolysis in inflamed adipocytes thus elevating serum FFA level which in turn adversely affects insulin sensitivity^{88–90}. Several recent reports emphasize the pathophysiological association between macrophages and adipose tissue inflammation where excess lipid plays the key role. The link between immunity and adipose tissue inflammation that cause T2D poses challenge for developing novel therapeutic interventions that can cover all these intricate complications involved in this insidious disease.

Conclusion

Research during the past few decades has identified adipose tissue as the dominant regulator of energy homeostasis. Its inflammation due to over-supply of lipid not only leads to insulin resistance where uptake and tolerance to glucose is attenuated but also adversely affects

lipid uptake, storage and mobilization. FetA is a novel linker between saturated fatty acid and TLR4. When FetA presents fatty acids to TLR4, it mediates inflammation through NFkB dependent pathway and starts secreting proinflammatory cytokines, TNF α and IL6, which in turn cause insulin resistance. This is further aggravated by the mobilization of macrophage into inflamed adipose tissue, which are transformed to pro-inflammatory M1 phenotype and release proinflammatory cytokines. These together lead to an intense inflammatory state which culminates in insulin resistance reaching a critical stage that leads to the onset of type 2 diabetes.

- Kahn, B. B., Type 2 diabetes: when insulin secretion fails to compensate for insulin resistance. *Cell*, 1998, 92, 593-596.
- Donath, M. Y., Targeting inflammation in the treatment of type 2 diabetes: time to start. Nat. Rev. Drug. Discov., 2014, 13, 465– 477
- Lackey, D. E. and Olefsky, J. M., Regulation of metabolism by the innate immune system. *Nat. Rev. Endocrinol.*, 2016, 12, 15–858.
- 4. Zimmet, P., Alberti, K. G. and Shaw, J., Global and societal implications of the diabetes epidemic. *Nature*, 2001, **13**, 782–787.
- Santomauro, A. T. et al., Overnight lowering of free fatty acids with acipimox improves insulin resistance and glucose tolerance in obese diabetic and nondiabetic subjects. Diabetes, 1999, 48, 1836–1841.
- Cusi, K., Kashyap, S., Gastaldelli, A., Bajaj, M. and Cersosimo, E., Effect on insulin secretion and insulin action of a 48-h reduction of plasma free fatty acids with acipimox in nondiabetic subjects genetically predisposed to type 2 diabetes. *Am. J. Physiol. Endocrinol. Metab.*, 2007, 292, 1775–1781.
- Boden, G., Obesity and free fatty acids (FFA). Endocrinol. Metab. Clin. North. Am., 2008, 37, 635–646.
- 8. Heilbronn, L. K. and Campbell, L. V., Adipose tissue macrophages, low grade inflammation and insulin resistance in human obesity. *Curr. Pharm. Des.*, 2008, **14**, 1225–1230.
- Lumeng, C. N. and Saltiel, A. R., Inflammatory links between obesity and metabolic disease. J. Clin. Invest., 2011, 121, 2111– 2117.
- Weisberg, S. P., McCann, D., Desai, M., Rosenbaum, M., Leibel, R. L. and Ferrante Jr, A. W., Obesity is associated with macrophage accumulation in adipose tissue. *J. Clin. Invest.*, 2003, 112, 1796–1808.
- 11. Xu, H. *et al.*, Chronic inflammation in fat plays a crucial role in the development of obesity-related insulin resistance. *J. Clin. Invest.*, 2003, **112**, 1821–1830.
- Glass, C. K. and Olefsky, J. M., Inflammation and lipid signalling in the etiology of insulin resistance. *Cell Metab.*, 2012, 15(5), 635-645.
- 13. McNelis, J. M. and Olefsky, J. M., Macrophages, immunity, and metabolic disease. *Cell (Immunity)*, 2014, **41**, 36–48.
- Nickerson, J. G. et al., Greater transport efficiencies of the membrane fatty acid transporters FAT/CD36 and FATP4 compared with FABPpm and FATP1 and differential effects on fatty acid esterification and oxidation in rat skeletal muscle. J. Biol. Chem., 2009, 284, 16522–16530.
- Watt, M. J. and Hoy, A. J., Lipid metabolism in skeletal muscle: generation of adaptive and maladaptive intracellular signals for cellular function. Am. J. Physiol. Endocrinol. Metab., 2012, 302, 1315-1328.
- Rosen, E. D. and Spiegelman, B. M., Adipocytes as regulators of energy balance and glucose homeostasis. *Nature*, 2006, 444(7121), 847–853.

- Guilherme, A., Virbasius, J. V., Puri, V. and Czech, M. P., Adipocyte dysfunctions linking obesity to insulin resistance and type 2 diabetes. *Nat. Rev. Mol. Cell Biol.*, 2008, 9(5), 367–377.
- Hajer, G. R., van Haeften, T. W. and Visseren, F. L., Adipose tissue dysfunction in obesity, diabetes, and vascular diseases. *Eur. Heart. J.*, 2008, 29, 2959–2971.
- Krotkiewski, M., Bjorntorp, P., Sjostrom, L. and Smith, U., Impact of obesity on metabolism in men and women. Importance of regional adipose tissue distribution. *J. Clin. Invest.*, 1983, 72, 1150–1162.
- Tchoukalova, Y. D., Votruba, S. B., Tchkonia, T., Giorgadze, N., Kirkland, J. L. and Jensen, M. D., Regional differences in cellular mechanisms of adipose tissue gain with overfeeding. *Proc. Natl. Acad. Sci. USA*, 2010, 107, 18226–18231.
- Tchkonia, T. et al., Fat tissue, aging, and cellular senescence. Aging Cell, 2010, 9, 667–684.
- Jensen, M. D., Haymond, M. W., Rizza, R. A., Cryer, P. E. and Miles, J. M., Influence of body fat distribution on free fatty acid metabolism in obesity. *J. Clin. Invest.*, 1989, 83, 1168–1173.
- 23. Boden, G., Obesity and free fatty acids. *Endocrinol. Metab. Clin. N. Am.*, 2008, **37**, 635–646.
- Halberg, N., Wernstedt-Asterholm, I. and Scherer, P. E., The adipocyte as an endocrine cell. *Endocrinol. Metab. Clin. N. Am.*, 2008, 37, 753–768.
- Stout, M. B. et al., Growth hormone action predicts age-related white adipose tissue dysfunction and senescent cell burden in mice. Aging, 2014, 6, 575-586.
- He, W. et al., Adipose-specific peroxisome proliferator-activated receptor gamma knockout causes insulin resistance in fat and liver but not in muscle. Proc. Natl. Acad. Sci. USA, 2003, 23, 100, 15712–15717.
- Choi, J. H. et al., Anti-diabetic drugs inhibit obesity-linked phosphorylation of PPAR gamma by Cdk5. Nature, 2010, 466, 451–456.
- Ahmadian, M., Myoung Suh, Jae, Hah, N., Liddle, C., Atkins, A. R., Downes, M. and Evans, R. M., PPARγ signaling and metabolism: the good, the bad and the future. *Nat. Med.*, 2013, 99, 557–566.
- 29. Tontonoz, P., Nagy, L., Alvarez, J. G. A., Thomazy, V. A. and Evans, R. M., PPARγ promotes monocyte/macrophage differentiation and uptake of oxidized LDL. *Cell*, 1998, **93**, 241–252.
- Berg, A. H., Combs, T. P. and Scherer, P. E. ACRP30/ adiponectin: an adipokine regulating glucose and lipid metabolism. *Trends Endocrinol. Metab.*, 2002, 13, 84–89.
- 31. Bonen, A. et al., Regulation of fatty acid transport by fatty acid translocase/CD36. Proc. Nutr. Soc., 2004, 63, 245–249.
- 32. Makowski, L. and Hotamisligil, G. S., Uncoupling of obesity from insulin resistance through a targeted mutation in aP2, the adipocyte fatty acid binding protein *J. Nutr.*, 2004, **134**, 2464–2468.
- 33. Boden, G. and Shulman, G. I., Free fatty acids in obesity and type 2 diabetes: defining their role in the development of insulin resistance and B-cell dysfunction. *Eur. J. Clin. Invest.*, 2002, 32, 14–23.
- Boden, G. and Chen, X., Effects of fat on glucose uptake and utilization in patients with non-insulin-dependent diabetes. J. Clin. Invest., 1995, 96, 1261–1268.
- 35. Boden, G., Chen. X., Rosner, J. and Barton, M., Effects of a 48-h fat infusion on insulin secretion and glucose utilization. *Diabetes*, 1995, 44, 1239–1242.
- Unger, R. H., Lipotoxicity in the pathogenesis of obesitydependent NIDDM. Genetic and clinical implications. *Diabetes*, 1995. 44, 863–870.
- Savage, D. B., Petersen, K. F. and Shulman, G. I., Disordered lipid metabolism and the pathogenesis of insulin resistance. *Physiol. Rev.*, 2007, 87, 507–520.
- 38. Donath, M. Y. and Shoelson, S. E., Type 2 diabetes as an inflammatory disease. *Nat. Rev. Immunol.*, 2011, **2**, 98–107.

- Braiman, L., Alt, A., Kuroki, T., Ohba, M., Bak, A., Tennenbaum, T. and Sampson, S. R., Insulin induces specific interaction between insulin receptor and protein kinase C delta in primary cultured skeletal muscle. *Mol. Endocrinol.*, 2001, 15, 565–574.
- Brunetti, A., Manfi, G., Chiefari, E., Goldfi, I. D. and Foti, D., Transcriptional regulation of human insulin receptor gene by the high-mobility group protein HMGI(Y); FASEB J., 2001, 15, 492– 500.
- Dey, D., Mukherjee, M., Basu, D., Datta, M., Roy, S. S., Bandyopadhyay, A. and Bhattacharya, S., Inhibition of insulin receptor gene expression and insulin signalling by fatty acid: interplay of PKC isoforms therein. *Cell. Physiol. Biochem.*, 2005, 16, 217–228.
- 42. Bhattacharya, S., Dey, D. and Roy, S. S., Molecular mechanism of insulin resistance. *J. Biosci.*, 2007, **32**, 405–413.
- Ruddock, M. W., Stein, A., Landaker, E., Park, J., Cooksey, R. C., McClain, D. and Patti, M. E., Saturated fatty acids inhibit hepatic insulin action by modulating insulin receptor expression and postreceptor signalling. *J. Biochem.*, 2008, 144, 599–607.
- Dasgupta, S., Bhattacharya, S., Maitra, S., Pal, D., Majumdar, S. S., Datta, A. and Bhattacharya, S., Mechanism of lipid-induced insulin resistance: activated PKCε is a key regulator. *Biochim. Biophys. Acta*, 2011, 1812, 495–506.
- 45. Nguyen, M. T. et al., A subpopulation of macrophages infiltrates hypertrophic adipose tissue and is activated by free fatty acids via Toll-like receptors 2 and 4 and JNK-dependent pathways. J. Biol. Chem., 2007, 282, 35279–35292.
- Yu, R., Kim, C. S., Kwon, B. S. and Kawada, T., Mesenteric adipose tissue-derived monocyte chemoattractant protein-1 plays a crucial role in adipose tissue macrophage migration and activation in obese mice. *Obesity*, 2006, 14, 1353–1362.
- 47. Johnson, A. M. F. and Olefsky, J. M., The origin and drivers of insulin resistance. *Cell*, 2013, **152**, 673–684.
- 48. Lee, J. Y., Sohn, K. H., Rhee, S. H. and Hwang, D., Saturated fatty acids, but not unsaturated fatty acids, induce the expression of cyclooxygenase-2 mediated through toll-like receptor 4. *J. Biol. Chem.*, 2001, 276, 16683–16689.
- Shi, H., Kokoeva, M. V., Inouye, K., Tzameli, I., Yin, H. and Flier, J. S., TLR4 links innate immunity and fatty acid-induced insulin resistance. *J. Clin. Invest.*, 2006, 116, 3015–3025.
- Schaeffler, A. et al., Fatty acid-induced induction of toll-like receptor-4/nuclear factor-kappaB pathway in adipocytes links nutritional signalling with innate immunity. *Immunology*, 2009, 126, 233–245.
- Saberi, M. et al., Hematopoietic cell-specific deletion of toll-like receptor 4 ameliorates hepatic and adipose tissue insulin resistance in high-fat-fed mice. Cell Metab., 2009, 10, 419–429.
- Kim, J. K., Fat uses a toll-road to connect inflammation and diabetes. Cell Metab., 2006, 4, 417–419.
- Kim, F. et al., Toll-like receptor-4 mediates vascular inflammation and insulin resistance in diet-induced obesity. Circ. Res., 2007, 100, 1589–1596.
- 54. Fessler, M. B., Rudel, L. L. and Brown, J. M., Toll-like receptor signaling links dietary fatty acids to the metabolic syndrome. *Curr. Opin. Lipidol.*, 2009, **20**, 379–385.
- Erridge, C. and Samani, N. J., Saturated fatty acids do not directly stimulate toll-like receptor signalling. *Arterioscler. Thromb. Vasc. Biol.*, 2009, 29, 1944–1949.
- Dasgupta, S., Bhattacharya, S., Biswas, A., Majumdar, S. S., Mukhopadhyay, S., Ray, S. and Bhattacharya, S., NF-kB mediates lipid-induced fetuin-A expression in hepatocytes that impairs adipocyte function effecting insulin resistance. *Biochem. J.*, 2010, 429, 451–462.
- Stefan, N. et al., α2-Heremans-Schmid glycoprotein/fetuin-A is associated with insulin resistance and fat accumulation in the liver in humans. Diabetes Care, 2006, 29, 853-857.
- Ix, J. H. et al., Fetuin-A and incident diabetes mellitus in older persons. J. Am. Med. Assoc., 2008, 300, 182–188.

- Brix, J. M., Sting, H., Höllerl, F., Schernthaner, G. H., Kopp, H.
 P. and Schernthaner, G., Elevated Fetuin-A concentrations in morbid obesity decrease after dramatic weight loss, *J. Clin. Endo*crinol. Metab., 2010, 95, 4877–4881.
- Stefan, N. and Häring, H. U., Circulating fetuin-A and free fatty acids interact to predict insulin resistance in humans. *Nat. Med.*, 2013, 19, 394–395.
- 61. Trepanowski, J. F., Mey, J. and Varady, K. A., Fetuin-A: a novel link between obesity and related complications. *Int. J. Obes.*, 2015, 39, 734–741.
- Pal, D. et al., Fetuin-A acts as an endogenous ligand of TLR4 to promote lipid-induced insulin resistance. Nat. Med., 2012, 18, 1279–1285.
- 63. Chatterjee, P. *et al.*, Adipocyte fetuin-A contributes to macrophage migration into adipose tissue and their polarization. *J. Biol. Chem.*, 2013, **288**, 28324–28330.
- Mathews, S. T. et al., Improved insulin sensitivity and resistance to weight gain in mice null for the Ahsg gene. *Diabetes*, 2002, 51, 2450-2458.
- 65. DeFronzo, R. A., Jacot, E., Jequier, E., Maeder, E., Wahren, J. and Felber, J. P., The effect of insulin on the disposal of intravenous glucose: results from indirect calorimetry and hepatic and femoral venous catheterization. *Diabetes*, 1981, 30, 1000–1007.
- 66. Shulman, G. I., Rothman, D. L., Jue, T., Stein, P., DeFronzo, R. A. and Shulman, R. G., Quantitation of muscle glycogen synthesis in normal subjects and subjects with non-insulin dependent diabetes by ¹³C nuclear magnetic resonance spectroscopy. N. Engl. J. Med., 1990, 322, 223–228.
- 67. Saltiel, A. R. and Kahn, C. R., Insulin signalling and the regulation of glucose and lipid metabolism. *Nature*, 2001, **414**, 799–806.
- 68. Fink, L. N. *et al.*, Pro-inflammatory macrophages increase in skeletal muscle of high fat-Fed mice and correlate with metabolic risk markers in humans. *Obesity (Silver Spring)*, 2014, **22**, 747–757.
- 69. Morino, K., Petersen, K. F. and Shulman, G. I., Molecular mechanisms of insulin resistance in humans and their potential links with mitochondrial dysfunction. *Diabetes*, 2006, **55**, 9–15.
- Schenk, S., Saberi, M. and Olefsky, J. M., Insulin sensitivity: modulation by nutrients and inflammation. J. Clin. Invest., 2008, 118, 2992–3002.
- 71. Houmard, J. A., Intramuscular lipid oxidation and obesity. *Am. J. Physiol. Regul. Integr. Comp. Physiol.*, 2008, **294**, 1111–1116.
- Lehr, S. et al., Identification and Validation of Novel Adipokines Released from Primary Human Adipocytes. Mol. Cell Proteomics, 2012, 11.
- 73. Breitling, R., Robust signaling networks of the adipose secretome. *Trends Endocrinol. Metab.*, 2009, **20**, 1–7.
- Samuel, V. T. and Shulman, G. I., Mechanisms for insulin resistance: common threads and missing links. *Cell*, 2012, 148, 852–871.
- 75. Jornayvaz, F. R. and Shulman, G. I., Diacylglycerol activation of protein kinase $C\varepsilon$ and hepatic insulin resistance. *Cell Metab.*, 2012. **15**. 574–584.
- Kelley, D. E, Mokan, M., Simoneau, J. A. and Mandarino, L. J., Interaction between glucose and free fatty acid metabolism in human skeletal muscle. J. Clin. Invest., 1993, 92, 91–98.
- Petersen, K. F., Dufour, S., Befroy, D., Garcia, R. and Shulman, G. I., Impaired mitochondrial activity in the insulin-resistant offspring of patients with type 2 diabetes. N. Engl. J. Med., 2004, 350, 664–671.

- Turner, N., Bruce, C. R., Beale, S. M., Hoehn, K. L., So, T., Rolph, M. S. and Cooney, G. J., Excess lipid availability increases mitochondrial fatty acid oxidative capacity in muscle: evidence against a role for reduced fatty acid oxidation in lipid-induced insulin resistance in rodents. *Diabetes*, 2007, 56, 2085– 2092
- Sparks, L. M., Xie, H., Koza, R. A., Mynatt, R., Hulver, M. W., Bray, G. A. and Smith, S. R., A high-fat diet coordinately downregulates genes required for mitochondrial oxidative phosphorylation in skeletal muscle. *Diabetes*, 2005, 54, 1926–1933.
- Brehm, A., Krssak, M., Schmid, A. I., Nowotny, Peter, Waldha, W. and Roden, M., Increased lipid availability impairs insulinstimulated ATP synthesis in human skeletal muscle. *Diabetes*, 2006, 55, 136–140.
- 81. Chanseaume, E. et al., Diets high in sugar, fat, and energy induce muscle type-specific adaptations in mitochondrial functions in rats. J. Nutr., 2006, 136(8), 2194–2200.
- 82. Koves, T. R. *et al.*, Mitochondrial overload and incomplete fatty acid oxidation contribute to skeletal muscle insulin resistance. *Cell Metab.*, 2008, 7, 45–56.
- 83. Muoio, D. M. and Neufer, P. D., Lipid-induced mitochondrial stress and insulin action in muscle. *Cell Metab.*, 2012, **15**, 595–605
- Wicks, S. E. et al., Impaired mitochondrial fat oxidation induces adaptive remodeling of muscle metabolism. Proc. Natl. Acad. Sci. USA, 2015, 112, 3300–3309.
- 85. Chen, A. *et al.*, Diet induction of monocyte chemoattractant protein-1 and its impact on obesity. *Obes. Res.*, 2005, **13**, 1311–1320
- Gutierrez, D. A., Kennedy, A., Orr, J. S., Anderson, E. K., Webb, C. D., Gerrald, W. K. and Hasty, A. H., Aberrant accumulation of undifferentiated myeloid cells in the adipose tissue of CCR2deficient mice delays improvements in insulin sensitivity. *Diabetes*, 2011, 60, 2820–2829.
- 87. Oh, D. Y., Morinaga, H., Talukdar, S., Bae, E. J. and Olefsky, J. M., Increased macrophage migration into adipose tissue in obese mice. *Diabetes*, 2012, **61**, 346–354.
- Hotamisligil, G. S., Arner, P., Caro, J. F., Atkinson, R. L. and Spiegelman, B. M., Increased adipose tissue expression of tumor necrosis factor-α in human obesity and insulin resistance. *J. Clin. Invest.*, 1995, 95, 2409–2415.
- Wellen, K. E. and Hotamisligil, G. S., Obesity-induced inflammatory changes in adipose tissue. *J. Clin. Invest.*, 2003, 112, 1785–1788.
- 90. Langin, D. and Arner, P., Importance of TNF α and neutral lipases in human adipose tissue lipolysis. *Trends Endocrinol. Metab.*, 2006, 17, 314–320.

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